

REVIEW

Interpreting Alzheimer's disease clinical trials in light of the effects on amyloid- β

Jeremy H Toyn and Michael K Ahljianian*

Abstract

The failure of several potential Alzheimer's disease therapeutics in mid- to late-stage clinical development has provoked significant discussion regarding the validity of the amyloid hypothesis. In this review, we propose a minimum criterion of 25% for amyloid- β ($A\beta$) lowering to achieve clinically meaningful slowing of disease progression. This criterion is based on genetic, risk factor, clinical and preclinical studies. We then compare this minimum criterion with the degree of $A\beta$ lowering produced by the potential therapies that have failed in clinical trials. If the proposed minimum $A\beta$ lowering criterion is used, then the amyloid hypothesis has yet to be adequately tested in the clinic. Therefore, we believe that the amyloid hypothesis remains valid and remains to be confirmed or refuted in future clinical trials.

Introduction and scope

Alzheimer's disease (AD) is a devastating and costly disease accounting for 50 to 80% of senile dementia cases. Worldwide, over 35 million people have dementia and the number is projected to double in the next 20 years [1]. Current treatments for symptoms have marginal benefits, and none treat the disease itself. A key hallmark of AD brain pathology is the accumulation of amyloid plaques. These consist largely of amyloid- β ($A\beta$) peptide, which is formed through proteolytic cleavage of amyloid precursor protein (APP) by two proteases: β -site APP-cleaving enzyme (BACE) and γ -secretase. Rare mutations in APP and the catalytic subunit of γ -secretase, presenilin, cause inherited forms of AD (familial AD (FAD)) with accelerated age of onset. In addition there are genetic risk factors, such as apoE4 and the APP Iceland mutant, that respectively increase or decrease AD risk. These genetic polymorphisms are all associated with changes in the production of $A\beta$, or changes in the relative amount of the more neurotoxic 42 amino acid form of $A\beta$, 'A β 42' [2]. Thus, genetic and pathological evidence has converged on the amyloid hypothesis of AD, proposing that accumulation of $A\beta$ is neurotoxic, leading to neuron loss, dementia and death [3,4]. Accordingly, major approaches to AD drug development

over the past two decades have focused on lowering $A\beta$ - for example, by inhibition of BACE or γ -secretase, or by the use of therapeutic antibodies to neutralize or enhance clearance of $A\beta$. Unfortunately, several clinical trials based on these approaches have been unsuccessful, raising the question of whether failure was due to insufficient target engagement, trial design, or the amyloid hypothesis. Here we address the target engagement question: what is the minimum extent of $A\beta$ lowering sufficient for significant cognitive benefit in AD patients? And has this level of target engagement yet been achieved in patients for sufficient trial duration?

Evidence in humans for the effect of changes in amyloid- β production

Human genetic evidence suggests that modest changes in $A\beta$ production are associated with a significant impact on AD. FAD mutants in which the APP gene is duplicated increase the gene dosage of APP by 50%, implying increased $A\beta$ production [5]. This suggests that a 33% decrease of $A\beta$ production in affected individuals would result in $A\beta$ production rates equivalent to that of normal healthy individuals. A similar situation of 50% increased APP gene dosage due to trisomy 21 is associated with >50% increase in APP mRNA expression, and may contribute to early onset AD in Down's syndrome [6]. In sporadic (late onset) AD, a 30% decreased clearance of $A\beta$ was reported in AD subjects, based on data using a heavy isotope labeling method [7]. In contrast to

* Correspondence: michael.ahljianian@bms.com
Bristol-Myers Squibb Research and Development, Neuroscience Biology, 5
Research Parkway, Wallingford, Connecticut 06492, USA

the FAD mutants, one rare APP mutant was associated with decreased incidence of AD [8]. In cell cultures overexpressing this mutant, BACE cleavage of the mutant APP was decreased by 50%, thereby decreasing A β production. This result implies that A β production in heterozygous individuals would be decreased by about 25%, although direct measurements of A β production in these individuals have not been reported. Thus, accumulating evidence suggests that relatively modest changes in A β , perhaps as little as 25% change over a sufficient period of time, can have a significant impact on AD.

In addition to the association of decreased A β levels with decreased disease risk, increased production of A β 42, relative to other A β peptides, is associated with earlier age of disease onset. Studies of A β production in cell cultures expressing presenilin FAD mutants showed that the relative amount of A β 42, measured as an A β 42/A β 40 ratio, was inversely correlated with age of onset [9,10]. To a first approximation, an earlier age of onset by 1 year was associated with a 1% increased A β 42/A β 40 production ratio, as measured in cell cultures. Another study reported an FAD mutant in which A β 40 was selectively decreased without change in A β 42, thus further emphasizing the role of the ratio [11]. A β 42/A β 40 production ratios are more challenging to measure *in vivo*, and require methods that circumvent the confounding effects of A β aggregation and degradation in the brain. The recent stable isotope labeling study by Potter *et al.* [12] reported that presenilin FAD subjects had a 24% increased A β 42/A β 40 production ratio, and selectively increased A β 42 clearance in subjects with amyloid deposits. This is consistent with the amyloid hypothesis; increased A β 42 production leads to increased aggregation in the brain, thereby decreasing the amount of A β 42 transported into the cerebrospinal fluid (CSF). This results in the counterintuitive situation in which increased A β 42 levels in brain lead to decreased A β 42 in CSF. In an earlier report using the stable isotope labeling method, sporadic AD patients (who were not FAD carriers) had decreased clearance in A β 42 and A β 40 of 30% and 26%, respectively, but no difference in production rates relative to age-matched controls [7]. Clearly, more studies are required to understand differences in A β dynamics between different genotypes and stages of disease, but thus far it appears that increases in either total A β or A β 42 production can accelerate disease onset. In contrast to presenilin FAD mutants, APP FAD mutants were reported to increase A β 38 production, in addition to A β 42, relative to other A β peptides, and *in vitro* results raised the possibility that A β 38 may also contribute to aggregation and neurotoxicity [13]. Thus, small changes, most likely less than 25%, in the ratios of A β peptides are associated with profound changes in AD risk and age of onset.

The human evidence described in the above section is summarized in Table 1.

Evidence from Alzheimer's disease mouse models for the effect of changes in amyloid- β levels on cognition

APP transgenic (TgAPP) mice are engineered to overexpress human APP, and in most cases exhibit A β -dependent pathology and cognitive deficits. Multiple genetic and pharmacological methods have been used to explore A β changes in these models. The soluble pool of A β responds rapidly to changes in A β production, whereas amyloid plaque-associated A β accumulates slowly with age, and does not respond acutely to changes in A β production. Therefore, we first considered studies that reported measurements of soluble A β -lowering and associated cognitive outcomes in TgAPP mice (Table 2).

BACE1 knock out (KO) mice exhibited a range of A β lowering from 12% for heterozygous to >90% for homozygous animals, with cognitive benefits in multiple types of cognitive assays [14-20]. In contrast, ablation of γ -secretase caused developmental abnormal or lethal phenotypes, and conditional KO (cKO) alleles of presenilin or nicastrin caused neurodegeneration and memory deficits in wild-type mice [21-24]. Thus, it is hardly surprising that presenilin cKO did not consistently show cognitive benefits in TgAPP mice despite A β lowering in the 55 to 75% range [25,26]. For γ -secretase ablation, it is possible that any benefit of A β lowering is confounded by deficits caused by loss of other functions of γ -secretase, such as Notch receptor activation. In addition, the restriction of the presenilin cKO allele to the forebrain may not have targeted A β lowering to the optimal anatomical location for benefit in TgAPP. A repressible TgAPP allele has been used to control A β synthesis in TgAPP mice [27]. In this study, aged plaque-bearing mice were fed doxycycline to repress TgAPP expression, implying a corresponding decrease in newly synthesized A β . Cognitive improvement was detected after 7 days, and yet no detectable lowering of transgene-derived soluble A β 42 was apparent, presumably due to equilibrium of soluble A β 42 with plaque A β 42. A β can also be decreased by cystatin C KO, which increases A β clearance via increased cathepsin B protease activity. Cognitive benefits in cystatin KO mice were associated with A β lowering of about 40% in young plaque-free TgAPP mice [28].

Improved cognition in TgAPP mice chronically dosed with BACE inhibitors (BACEis) GRL-8234, TAK-070 and trihydroxychalcone was associated with amyloid plaque lowering in the 20 to 60% range, but no evidence of decreased A β production was reported [29-31]. TgAPP mice given single doses of the γ -secretase inhibitors (GSIs) DAPT, begacestat, semagacestat, and avagacestat

Table 1 Alzheimer's disease and human A β levels

Disease state	Affect on A β	Type of study	Reference
APP gene duplication	50% \uparrow production inferred	Gene copy number in patients	[5]
Trisomy 21	50% \uparrow production inferred	mRNA levels in patients	[6]
Protective APP allele	25% \downarrow production	Cell culture	[8]
Presenilin FAD	A β 42/A β 40 1% \uparrow per year earlier age of onset	Cell culture	[9,10]
Presenilin FAD	A β 42/A β 40 24% \uparrow clearance	Patient CSF samples	[12]
Sporadic AD	A β 42 30% \downarrow clearance; A β 42 26% \downarrow clearance	Patient CSF samples	[7]
APP FAD	A β 42 \uparrow and A β 38 \uparrow production	Cell culture	[13]

A β , amyloid- β ; AD, Alzheimer's disease; APP, amyloid precursor protein; CSF, cerebrospinal fluid; FAD, familial Alzheimer's disease.

showed cognitive improvements with A β lowering in the range 0 to 35% [32-36]. The effect of a single dose is noteworthy because it implies an acute role of newly synthesized A β in cognitive impairment. In the study by Mitani *et al.* [35], a 1 mg/kg single dose of semagacestat or avagacestat improved Y maze performance, although decreased A β was only detectable at higher doses. However, 8-day repeat dosing at 1 mg/kg did not improve Y maze performance in TgAPP mice, and actually impaired Y maze performance in wild-type mice. Thus, like the presenilin cKO allele, it appears any benefit of A β lowering in TgAPP mice may have been confounded by other deficits resulting from γ -secretase inhibition, in this case

proposed due to accumulation of APP β -CTF fragment [35].

Selective lowering of A β 42 is of therapeutic interest because of increased A β 42 in FAD mutants, the evidence that A β 42 is the earliest deposited species [37], and the cognitive disruption caused by A β 42 aggregates in animal models [38,39]. Furthermore, *in vitro* studies have shown that A β 42 aggregation is inhibited by A β 40 [40-42], and also by A β 37 and A β 38 [43], suggesting that the shorter peptides are capable of interfering with the amyloid cascade. A variety of genetic and pharmacological methods have been used to selectively alter A β 42 levels *in vivo*. An increased A β 42/A β 40 ratio enhanced

Table 2 A β -lowering cognitive benefit in TgAPP mice

A β -lowering method	Brain A β lowering (%)	Observed functional benefits	Mouse strain	Reference
BACE1 $-/-$ KO	>90%	Contextual fear conditioning, Morris water maze, social recognition	Tg2576; 5x FAD	[14-16]
BACE+/- KO	12% in young mice	Contextual fear conditioning, conditioned taste aversion	PDAPP; 5x FAD	[17-20]
Presenilin conditional forebrain KO	75%	No benefit (novel object recognition in 3- to 6-month-old mice)	APP [V717I]	[25]
Presenilin conditional forebrain KO	55% in young mice	Contextual fear conditioning and Morris water maze in young but not old mice	APP J20	[26]
TgAPP conditional allele	\geq 70% new A β ; no effect on steady-state A β 42 levels	Two trial Y maze; plus water maze; radial arm water maze	Repressible TgAPP	[27]
Cystatin C KO	40% (A β); 60% (A β 42) in young mice	Morris water maze	APP J20	[28]
GRL-8234 (BACEi)	35-50% plaque after 7 months	Morris water maze	Tg2576	[31]
TAK-070 (BACEi)	20% plaque after 7 weeks	Y-maze, Morris water maze, novel object recognition	Tg2576	[29]
Trihydroxychalcone (BACEi)	50-60% plaque after 106 days	Morris water maze	APP-PS1	[28]
DAPT (GSI) single dose	25% at 8 hours	Contextual fear conditioning	Tg2576	[32]
DAPT (GSI) repeat dose	35% after 4 days	Morris water maze	Ts65Dn	[34]
Begacestat (GSI) single dose	25-35% at 4 hours	Contextual fear conditioning	Tg2576	[33]
Semagacestat/LY450139 (GSI)	No change 1 mg/kg; 25-30% 10 mg/kg	Y maze benefit at 1 mg/kg after single dose; no benefit at 10 mg/kg or 8-day repeat dosing	Tg2576	[35]
Avagacestat/ BMS-708163 (GSI)	No change 1 mg/kg; 25-30% 10 mg/kg	Y maze benefit at 1 mg/kg after single dose; no benefit at 10 mg/kg or 8-day repeat dosing	Tg2576	[35]

A β , amyloid- β ; BACE, β -site APP-cleaving enzyme; BACEi, β -site APP-cleaving enzyme inhibitor; GSI, γ -secretase inhibitor; KO, knock out; TgAPP, APP transgenic.

aggregation and neurotoxicity *in vitro* and caused memory deficits after a single intraventricular injection in wild-type mice [44]. A presenilin mutant that selectively lowered A β 40 exacerbated plaque deposition in TgAPP mice, implicating the A β 42/A β 40 ratio *per se* [45]. In another *in vivo* approach, novel Tg-A β 42 and Tg-A β 40 transgenes were used for selective expression of A β 42 or A β 40, respectively. Selective expression of A β 40 was shown to interfere with A β plaque accumulation in Tg-A β 42 and TgAPP mice [46]. Remarkably, however, Tg-A β 42 and Tg-A β 40 mice exhibited no cognitive defects in a range of tests, indicating that overexpression of A β was insufficient for neurotoxicity in this model [47]. As mentioned above, a cystatin C KO in TgAPP mice ameliorated cognition associated with 40% overall lowering of A β peptides; however, this was also in the context of 33% relative lowering of A β 42 [28].

γ -Secretase modulators (GSMs) include a variety of small molecules that target γ -secretase, causing decreased A β 42 and increased production of one or more shorter peptides such as A β 37, -38, or -39 [48]. Thus, GSMs have an essentially opposite effect to FAD mutants. The GSM EVP-0015962 improved cognition in TgAPP mice after a single dose that caused a 50% decrease in A β 42 [49]. CHF5074 improved cognition after chronic dosing in TgAPP mice with no discernable A β 42 lowering, but it seems probable that the cognitive effect was not related to the GSM activity of this compound, which is of very low potency [50-53]. TgAPP mice given single doses or 8-day repeat doses of GSM-2 at ≥ 0.1 mg/kg showed improved Y maze performance [35], although A β 42 lowering, of 20% and 30%, was detected only at the higher doses of 1 and 3 mg/kg, respectively [35,36].

The GSMs JNJ40418677 and 'compound 4' exhibited A β 42 lowering activity in the 40 to 50% range, but cognitive effects were not reported. However, long term dosing of these compounds did decrease A β plaque accumulation [54,55]. Thus, accumulating evidence suggests that decreased A β 42 relative to shorter A β production affects the amyloid cascade and improves cognitive performance in TgAPP models, as summarized in Table 3.

The interpretation of the evidence linking A β lowering and cognitive benefits in animal models should take several factors into account, including the mechanism by which A β lowering was achieved, and the possibility of confounding toxicity, as well as the observed change in A β levels. For example, sustained A β lowering is likely to be more impactful than transient A β lowering. For genetic methods of A β ablation, measurement of soluble A β levels at a single time point represents the overall sustained level of A β lowering. For small molecules, however, A β lowering data often refer to a single optimal time point after dosing, which can be several-fold greater than the average extent of A β lowering across the dosing interval. In addition, the form of A β measured should be considered. Many studies, including immunization approaches, have reported cognitive benefits associated with decreased plaque A β . Decreased plaque A β is a downstream endpoint, and is not a direct readout for decreased A β production or neurotoxic forms of A β . Nevertheless, such studies give further evidence of the link between the amyloid cascade and cognition [56].

Thus, taking into consideration a wide range of studies in TgAPP mice and human genetics, relatively modest

Table 3 γ -Secretase modulators and selective A β 42-lowering benefit

Selective A β 42-lowering method	Brain A β 42 lowering (%)	Observed cognitive or pathological benefits in deficient Tg mouse	Mouse strain	Reference
ICV injection of preaggregated A β 42/A β 40	A β 42/40 3:7 ratio; 1:9 ratio inactive	Passive avoidance and contextual fear conditioning	Wild type; intraventricular A β administration	[44]
BRI-A β 40 and BRI-A β 42 transgenes	50-400% increased A β 40 (decreased 42/total ratio)	60-90% decreased plaque; improved survival; however, these mice exhibited no A β -dependent cognitive phenotypes	Tg2576 and Tg-A β 40	[46,47]
EVP-0015962	50% after single 30 mpk dose	Contextual fear conditioning, gliosis 75% plaque load, after 50 weeks at 60 mpk/day	Tg2576	[49]
CHF5074	No significant change (4-9 month treatment)	Contextual memory, 50-75% decreased plaque burden, astrogliosis, synaptophysin levels, neurogenesis	Tg2576	[50-53]
GSM-2	0-30% at 0.1-3 mpk, respectively	Y maze improvements at 0.1-3 mpk in mice aged 5.5 months	Tg2576	[35]
GSM-2	50-60% nascent A β 2 hours after 10 mpk	Y maze and plaque pathology in mice aged 10-18 months	Tg2576	[36]
JNJ40418677	50% max lowering 30 mpk single dose	Up to 96% decreased plaque area and number after 7 months at 120 mpk/day	Tg2576	[55]
Compound 4	40% decrease 100 mpk single dose	48-76% decrease of plaque A β after 7 months at 50 mpk/day	Tg2576	[54]

A β , amyloid- β ; ICV, intracerebroventricular; mpk, mg/kg; Tg, transgenic.

decreases in A β , of about 25%, are associated with cognitive benefits (Tables 2 and 3). Therefore, we propose that sustained A β lowering of 25% using any method tolerated for a sufficient period of time in patients represents a reasonable minimal objective. While this criterion is proposed as a minimal objective, an optimal therapeutic will provide the flexibility to probe a range of A β -lowering activity, including nearly complete lowering, in order to understand the relationship between A β lowering and efficacy. Nevertheless, greater than 25% may not be achievable by some compounds, and consequently setting the bar too high could result in lost opportunities. Lowering of A β by approximately 25% therefore sets a reasonable starting point for the minimum level of pharmacodynamic effect to justify efficacy trials in AD patients.

Demonstration of amyloid- β lowering in recent clinical trials

If the preceding arguments are valid, then a pressing question is whether the recent, late stage clinical studies achieved the 25% A β lowering criterion. Before this question can be addressed, however, two antecedent questions require clarification; at what stage of AD might 25% A β reduction produce efficacy, and what form of A β should be targeted for 25% reduction.

What is the relationship between the extent of A β lowering required for efficacy and disease stage? For example, does it escalate with disease progression - that is, is the requirement for A β reduction lower if intervention is earlier (predementia/presymptomatic), and greater if intervention is later in disease (mild to moderate)? Alternatively, is there some degree of A β lowering that will produce efficacy regardless of stage of intervention? Finally, is there a point in the disease process that is unresponsive to A β -directed therapies (for example, moderate to severe)? While clear answers to these questions will not be forthcoming until an efficacious agent is identified, there seems to be consensus in the field that earlier intervention is desirable [4,57]. This consensus is based on the long latency of measurable pathologic changes (changes in CSF A β and tau, plaque and tangle development, volumetric magnetic resonance imaging (MRI)) and the relatively late onset of cognitive symptoms [58-64]. Based on the hypothesis that earlier intervention is better, several clinical efficacy studies targeting pre-symptomatic AD patients are either underway or planned (for example, [65,66]). For the purposes of this review, we propose the minimum criterion of 25% A β lowering for clinical trials targeting early stages of the disease, namely predementia (mild cognitive impairment with biomarker evidence consistent with AD) and mild AD. The combination of cognitive symptoms with biomarkers such as CSF A β 42, tau, volumetric MRI and

amyloid positron emission tomography (PET) suggest that these are the earliest disease stages for which a diagnosis of AD or likely progression to AD can currently be confidently assigned (for example, [59,67,68]). However, even mild AD may be too late for initiating A β -lowering therapies given the latency between biomarker positivity and symptom onset. Therefore, the 25% criterion could also be considered when designing trials for presymptomatic AD.

Which form of A β should be targeted for 25% reduction in efficacy trials? The amyloid hypothesis currently states that soluble A β is the species most deleterious to neuronal viability and synaptic function. While the precise molecular identity of the most toxic A β species is debatable (for example, [69]), the number of independently reproduced reports implicating soluble A β as disruptive to normal function strongly suggests that this species plays a key role in the cognitive decline observed in AD.

If soluble A β is the key culprit in cognitive impairment, how can sponsors assess potential reduction of this species in humans in clinical trials? Currently the best reflection of soluble brain A β is CSF A β [70]. CSF A β is used to aid in the diagnosis of AD [71-73] and has been used as a target engagement biomarker by sponsors developing therapies that are intended to lower A β [74,75]. The latter studies are typically supported by substantial preclinical data sets demonstrating an understanding of the relationship between brain and CSF A β -lowering produced by an A β -targeting compound in more than one species. These preclinical studies have demonstrated close correspondence between brain and CSF lowering of A β produced by GSIs [33,76-79], GSMs [70,79,80] and BACEis [70,81-83] confirming that CSF A β can reflect brain A β . These preclinical data sets are subsequently used as the basis for pharmacokinetic/pharmacodynamic (PK/PD) modeling to aid in dose selection and for determining the time points to sample CSF in human studies. For example, the GSI avagacestat produced reductions in rat brain A β that were reflected by comparable reductions in CSF after acute administration [78]. Modeling of these data accurately predicted the human PK/PD relationship for reductions in normal healthy volunteer (NHV) CSF [78,84,85]. Furthermore these PK/PD relationships for A β lowering did not differ significantly between NHVs and AD patients [86]. Additional preclinical data sets followed by PK/PD modeling and data collection in humans have been reported for other classes of A β -lowering drugs, including GSMs (BMS, unpublished) and BACEis [82,83]. Thus, for all synthesis-inhibitor mechanisms studied in this way, there is substantial correlation between lowering of A β in brain and CSF in preclinical species. Furthermore, modeling the preclinical data for translation has

faithfully predicted the PK/PD of CSF A β lowering in both NHVs and AD patients.

Nevertheless, the presence of plaques in patients presents a potential confound for interpreting or expecting changes in CSF A β in patients. There is still active debate regarding the role of amyloid plaques in producing the cognitive deficits observed in AD (for example, [87]). Neurons that are proximal to plaques display aberrant dystrophic neurites with disrupted trajectories indicative of synaptic dysfunction (for example, [88]) and plaques have been hypothesized to create and sustain neurotoxic microenvironments [89] and perturb mitochondrial function [90]. In patients, alterations in functional brain connectivity have been reported in plaque-bearing regions in cognitively normal subjects (for example, [91,92]). However, it remains unclear to what extent such proximal, plaque-associated dysfunction contributes to the global cognitive impairment observed in AD patients, particularly since cognitive function did not improve in a small number of patients with reduced plaque after treatment with AN1792 [93,94].

It has also been hypothesized that amyloid plaques are protective and serve as a mechanism for clearance of soluble amyloid species from the interstitial space (for example, [87]). Furthermore, under conditions in which soluble A β is decreased (for example, in the presence of a therapeutic that lowers soluble A β) there is speculation that the most recently plaque-associated A β may dissociate, re-attaining a soluble state in parenchyma and the potential to become toxic to neurons. Pre-clinically, measurements of interstitial A β suggest equilibrium between soluble and insoluble forms of A β [95] and a study that compared plaque removal by two antibodies that recognized either soluble or fibrillar A β demonstrated that only the fibril-preferring antibody decreased plaque load [96]. Finally, a recent study suggests that, in the presence of very low plasma A β , plaque volume does not change in Tg mice [97].

In the clinic, an examination of the PK/PD analyses comparing the CSF A β -lowering effects of avagacestat in AD patients and NHVs demonstrates little difference in these two populations, suggesting that the potential contribution of soluble A β derived from plaque may be modest [79,86]. Similarly, recent evidence from BACEi studies in AD patients and NHVs suggests that the potency for reducing A β peptides is equivalent in these human populations and that the fraction of CSF A β peptides that is not sensitive to BACE inhibition (and therefore may be derived from an alternative source, such as plaques) is quite small, ranging from 2 to 6% [98]. Furthermore, any association of soluble A β to plaques does not limit the ability to detect therapy-induced decreases in CSF A β in patients [86,99]. Thus, our view is that while soluble A β is likely to be in equilibrium

with plaques [12,95], and some fraction of soluble A β will associate with plaques, the data reported to date suggest that plaques are unlikely to provide a significant supply of soluble A β to CSF. Therefore, the ability to detect A β lowering in the CSF of AD patients should not be confounded by the presence of plaques, especially if lowering has been demonstrated in healthy volunteers. Nevertheless, the A β PET ligands have a clear role in AD diagnosis and can be used as target engagement biomarkers for some potential therapeutics, including antibodies [100,101].

How have the different A β -lowering mechanisms fared in late stage clinical trials? A summary is provided in Table 4. Avagacestat was tested in both mild-moderate and pre-dementia patient populations without evidence of efficacy, but the pre-dementia study was discontinued prior to the planned completion. The acute and steady state lowering of CSF A β produced by avagacestat in NHVs was substantial, but tolerability declined at doses that lowered CSF A β by more than approximately 15% in NHVs and especially in AD patients [85,86]. Thus, the maximum tolerated A β lowering was less than the 25% minimum criteria proposed above.

None of the published clinical data for semagacestat disclose evidence for steady state lowering of CSF A β in either NHVs [102] or AD patients [103,104]. However, a stable isotope labeling kinetic (SILK) study did demonstrate acute inhibition of the appearance of newly synthesized A β in the CSF [105] and reduction of shorter forms of A β have been interpreted as target engagement [106,107]. While the SILK study provided evidence for target engagement and inhibition of A β synthesis for a short period of time after dosing, the lack of steady state lowering of CSF A β at tolerated doses suggests that semagacestat may not have lowered soluble brain A β to a significant degree in NHVs or AD patients. Phase III studies demonstrated that semagacestat was not efficacious but exacerbated the cognitive decline in treated patients [108]. Thus, for the two GSIs that have achieved late stage clinical development, neither have achieved 25% lowering of soluble CSF A β in AD patients at tolerable doses and both have failed in the clinic. Taken together, the avagacestat and semagacestat examples suggest that, compared with NHVs, AD patients may be more sensitive to any unintended effects of potential therapeutics.

A small number of GSIs have also been tested in both NHVs and AD patients. Tarenflurbil failed in phase III [109], but CSF A β lowering was not reported in humans and the ability of the compound to lower brain A β in preclinical species has been the subject of debate [110,111]. CHF-5074 lowers A β in mouse models of APP overexpression but only after chronic dosing (that is, there are no sub-acute effects of this compound on

Table 4 Cerebrospinal fluid A β lowering - summary of clinical trial results

Name	Mechanism	Stage of development	NHVs	Patient population	Patient CSF A β ^a	Amyloid PET	Reference
AN1792	Active vaccine	D/C (phase IIa)	NR	M-M	No change	NR	[115]
Bapineuzumab	Passive vaccine	D/C (phase III (i.v.); phase II (s.c.))	NR	M-M	No change	Decrease	[74,116]
Solanezumab	Passive vaccine	Phase III, pre-sym	NR	M-M, mild	Total (40/42) - increased Unbound 42 - increased Unbound 40 - decreased	NR	[117,122]
Crenezumab	Passive vaccine	Phase I/II	NR	Pre-sym, FAD	NR	NR	[127]
Gantenerumab	Passive vaccine	Phase II/III	NR	M-M, pre-dem	NR	Decrease	[101]
IVIg	Anti-inflammatory	Phase III	NR	M-M	No change	NR	[126]
Tarenflurbil	GSM	D/C (phase III)	NR	M-M	NR	NR	[109]
Semagacestat	GSI	D/C (phase III)	No change	M-M	No change	NR	[102-104]
Avagacestat	GSI	D/C (phase II)	≥50% decrease	M-M	High dose: ~50% decrease Tolerated doses: ≤15% decrease	NR	[79,85,86]
LY2811376, LY2886721	BACE inhibitor	D/C (phase II)	≥50% decrease	M-M	NR	NR	[82]
MK8931	BACE inhibitor	Phase II	NR	M-M	≥80% decrease	NR	[99]

^aMost advanced stage clinical trial. A β , amyloid- β ; BACE, β -site APP-cleaving enzyme; CSF, cerebrospinal fluid; D/C, clinical development discontinued; FAD, familial Alzheimer's disease; GSI, γ -secretase inhibitor; GSM, γ -secretase modulator; i.v., intraventricular; M-M, mild to moderate Alzheimer's disease; NHV, normal healthy volunteer; NR, not reported; PET, positron emission tomography; pre-dem, predementia; pre-sym, presymptomatic; s.c., subcutaneous.

brain A β [51]) making PK/PD analyses challenging and calling into question the mechanism of A β lowering after chronic treatment. Nevertheless, this molecule has completed a small, 12-week phase II study in AD patients [112] and, while CSF A β was measured, no changes were reported. Several companies [113] have disclosed preclinical A β lowering data on GSMs and some of these publications include measurement of both brain and CSF and PK/PD analyses [80,81]. However, no clinical data have been released despite the disclosure of phase I studies sponsored by BMS (New York, NY, USA) and Eisai (Tokyo, Japan).

While clinical data for GSMs are scarce, there are excellent examples of preclinical PK/PD data sets generated with BACEis, with subsequent translation to clinical studies in one instance thus far [82,83]. Lilly (Indianapolis, IN, USA) has disclosed the most data, establishing a convincing relationship between brain and CSF lowering with subsequent PK/PD modeling and translation to humans [82]. Merck (Whitehouse Station, NJ, USA) has also disclosed clinical data with a BACEi [99]. The extent of lowering of CSF A β produced by both of these BACEis in NHVs and AD patients is unprecedented and can exceed 90%, suggesting that lowering of A β in the brain is substantial. Unfortunately, the lead Lilly molecule, LY2886721, produced hepatic adverse effects in AD patients (13 June 2013, Lilly press release) which

forced termination of the phase II study. However, the phase II development of the Merck BACEi continues, suggesting that the hepatic issues produced by LY2886721 may be off-target, compound-specific and unrelated to BACE inhibition. Importantly, these data indicate that BACE inhibition is currently the most promising therapeutic modality to directly test the A β hypothesis of AD.

Anti-amyloid- β antibodies and IVIG

More than a decade after the initial reports of positive effects on pathology and cognition produced by A β immunization in TgAPP mice [114], it is now well established that reduction of plaque volume and restoration of functional deficits in TgAPP mice can be achieved by both passive and active A β immunotherapy [56]. It was these findings in preclinical models that prompted clinical development of A β immunotherapy. However, the results from the late phase clinical studies assessing this modality have been predominantly negative. AN1792, an active vaccine, was discontinued in phase II due to meningoencephalitis [115] and produced a small increase in CSF A β . Two passive anti-A β immunotherapies, bapineuzumab and solanezumab, have completed phase III clinical studies. Intravenous administration of bapineuzumab failed [116], but a subcutaneous study continues. Solanezumab has completed two phase III studies with mixed results [117].

However, the data for solanezumab in mild AD patients were sufficiently encouraging to warrant an additional phase III study (3 July 2013, Eli Lilly press release).

What is the relationship of these results to the proposal that a minimum of 25% soluble A β reduction must be achieved to produce efficacy? Unfortunately, the answer to this question is unclear. Assessing the PK/PD of antibody therapy is more challenging than for small molecule therapy for several reasons, including antibodies that recognize multiple forms of A β (soluble, fibrillar), no direct measure of target engagement in brain (antigen-antibody complex), difficulty in assessing free antibody concentrations compared with total concentrations (that is, antibody bound to antigen), and a large pool of antibody in plasma that can exchange with brain antibody [118-120]. Preclinical studies assessing the effects of antibody therapy on CSF concentrations of A β are rare if not non-existent, leaving clinicians with very little data upon which to base the dose selection and dosing intervals required for efficacy. Nevertheless, in clinical studies for both solanezumab and bapineuzumab, assessments of CSF A β and tau were made. In the bapineuzumab phase II and III studies, no changes in CSF A β were detected and small decreases in both phospho-tau181 and total tau were reported [71,72,116,121]. For solanezumab the picture is more complex. In the phase II study, total CSF A β 40 and A β 42 increased. Unbound A β 42 also increased while unbound A β 40 was non-significantly decreased [122]. The increase in total CSF A β was interpreted as evidence for central nervous system penetration of the antibody while the increase in CSF A β 42 was suggested to be due to potential dissolution of amyloid plaques (see below). In the phase III studies, unbound A β 40 decreased while unbound A β 42 did not change compared to controls [117]. The increases in total CSF A β were interpreted as evidence for central nervous system penetration of the antibody while the changes in unbound A β peptides were suggested to be due to potential alterations in compartment equilibria (for example, central to peripheral or fibrillar to soluble). In summary, the A β immunotherapy data disclosed to date does not provide a clear picture on the effects on unbound, soluble CSF A β , suggesting that the utility of CSF A β as a target engagement biomarker for immunotherapy may be limited. Alternatively, the potential efficacy and biomarker effects of the antibodies may not have manifested due to limitations in dosing levels or frequency and the failure to achieve efficacious brain concentrations. For example, in contrast to small molecule therapy, the implications of these negative findings are difficult to interpret due to the lack of preclinical analyses that define a relationship between antibody exposure, brain and CSF A β , and functional measures of efficacy such as synaptic and cognitive measures.

IVIG is a purified preparation of human immunoglobulins that has been used therapeutically for immunodeficiency disorders. Based on preclinical data and the hypothesis that IVIG would provide a source of anti-A β antibodies and possibly anti-inflammatory activity [123,124], IVIG has been evaluated through phase III clinical trials. While reductions in total CSF A β were reported for small pilot studies [125], larger phase II studies resulted in no detectable changes in CSF A β [126]. The recently disclosed phase III study results demonstrated no treatment effect for IVIG (Baxter press release 7 May 2103).

A more commonly used measure of target engagement in clinical studies employing immunotherapy, especially for those antibodies that recognize fibrillar A β , is amyloid PET imaging (for example, [102,103]). As with CSF, however, very few, if any, preclinical analyses describe a relationship between plaque reduction and functional efficacy. Any such analysis would then require overlay or inclusion of immunotherapy PK to be helpful in dose selection for clinical trials.

Conclusion

The genetics and preclinical literature support the hypothesis that a 25% reduction in soluble A β is a scientifically based minimal criterion for any therapeutic directed toward lowering a soluble, pathologically relevant species of A β . Preclinical data demonstrate that soluble CSF A β can reflect soluble brain A β and PK/PD analyses of preclinical data reliably translate to the clinic for lowering of soluble CSF A β and, by inference, brain A β . While amyloid PET ligands can provide information on target engagement, especially for some antibodies, the relationship between plaque reduction, antibody exposure and efficacy has yet to be reported for any potential antibody therapeutic. The data from clinical trials disclosed to date suggest that no potential therapeutic has lowered soluble A β by 25%. Thus, while enormous progress has been made in understanding the basic mechanisms of AD and the identification of rational therapeutic mechanisms such as antibodies, GSIs, GSMs and BACEis, the amyloid hypothesis has yet to be adequately tested clinically by any of the current therapeutic moieties. Furthermore, the notion that the amyloid hypothesis is incorrect or has been disproven is premature. The potential of the current cohort of 'second generation' therapeutics, such as BACEis, which appear to provide potential for testing a broad range of A β lowering, and antibodies like crenezumab [127] and gantenerumab [128], is promising and may ultimately enable testing of the amyloid hypothesis.

Abbreviations

A β : Amyloid- β ; AD: Alzheimer's disease; APP: Amyloid precursor protein; BACE: β -site APP-cleaving enzyme; BACEi: β -site APP cleaving enzyme

inhibitor; cKO: Conditional knock out; CSF: Cerebrospinal fluid; FAD: Familial Alzheimer's disease; GSI: γ -secretase inhibitor; GSM: γ -secretase modulator; KO: Knock out; MRI: Magnetic resonance imaging; NHV: Normal healthy volunteer; PD: Pharmacodynamic; PET: Positron emission tomography; PK: Pharmacokinetic; SILK: Stable isotope labeling kinetic; TgAPP: APP transgenic.

Competing interests

The authors are employees of Bristol-Myers Squibb.

Acknowledgements

We thank our colleague Charlie Albright for critical review and suggestions during preparation of the manuscript.

Published: 12 Mar 2014

References

1. Wimo A, Prince M: *World Alzheimer Report 2010: the Global Economic Impact of Dementia*. [http://www.alz.org/documents/national/world_alzheimer_report_2010.pdf]
2. Findeis MA: **The role of amyloid β peptide 42 in Alzheimer's disease**. *Pharmacol Therapeut* 2007, **116**:266–286.
3. Hardy J, Selkoe DJ: **The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics**. *Science* 2002, **297**:353–356.
4. Karran E, Mercken M, De Strooper B: **The amyloid cascade hypothesis for Alzheimer's disease: an appraisal for the development of therapeutics**. *Nat Rev Drug Disc* 2011, **10**:698–712.
5. Rovelet-Lecruex A, Hannequin D, Raux G, Le Meur N, Laquerrière A, Vital A, Dumanchin C, Feuillette S, Brice A, Vercelletto M, Dubas F, Frebourg T, Campion D: **APP locus duplication causes autosomal dominant early-onset Alzheimer disease with cerebral amyloid angiopathy**. *Nat Genet* 2006, **38**:24–26.
6. Oyama F, Cairns NJ, Shimada H, Oyama R, Titani K, Ihara Y: **Down's syndrome: up-regulation of β -amyloid protein precursor and τ mRNAs and their defective coordination**. *J Neurochem* 1994, **62**:1062–1066.
7. Mawuenyega KG, Sigurdson W, Ovod V, Munsell L, Kasten T, Morris JC, Yarasheski KE, Bateman RJ: **Decreased clearance of CNS β -amyloid in Alzheimer's disease**. *Science* 2010, **330**:1774.
8. Jonsson T, Atwal JK, Steinberg S, Snaedal J, Jonsson PV, Bjornsson S, Stefansson H, Sulem P, Gudbjartsson D, Maloney J, Hoyte K, Gustafson A, Liu Y, Lu Y, Bhangale T, Graham RR, Huttenlocher J, Bjornsdottir G, Andreassen OA, Jönsson EG, Palotie A, Behrens TW, Magnusson OT, Kong A, Thorsteinsdottir U, Watts RJ, Stefansson K: **A mutation in APP protects against Alzheimer's disease and age-related cognitive decline**. *Nature* 2012, **488**:96–99.
9. Duering M, Grimm MO, Grimm HS, Schröder J, Hartmann T: **Mean age-of-onset in familial Alzheimer's disease is determined by amyloid beta 42**. *Neurobiol Aging* 2005, **26**:785–788.
10. Kumar-Singh S, Theuns J, Van Broeck B, Pirici D, Vennekens K, Corsmit E, Cruts M, Dermaut B, Wang R, Van Broeckhoven C: **Mean age-of-onset of familial Alzheimer disease caused by presenilin mutations correlates with both increased A β 42 and decreased A β 40**. *Hum Mutat* 2006, **27**:686–695.
11. Shimojo M, Sahara N, Murayama M, Ichinose H, Takashima A: **Decreased A β secretase by cells expressing familial Alzheimer's disease-linked mutant presenilin-1**. *Neurosci Res* 2007, **57**:446–453.
12. Potter R, Patterson BW, Elbert DL, Ovod V, Kasten T, Sigurdson W, Mawuenyega K, Blazey T, Goate A, Chott R, Yarasheski KE, Holtzman DM, Morris JC, Benzinger TL, Bateman RJ: **Increased *in vivo* amyloid- β 42 production, exchange and loss in presenilin mutation carriers**. *Sci Transl Med* 2013, **5**:189ra77.
13. Chávez-Gutiérrez L, Bammens L, Benilova I, Vandersteen A, Benurwar M, Borgers M, Lismont S, Zhou L, Van Cleynenbreugel S, Esselmann H, Wiltfang J, Serneels L, Karran E, Gijzen H, Schymkowitz J, Rousseau F, Broersen K, De Strooper B: **The mechanism of γ -secretase dysfunction in Alzheimer's disease**. *EMBO J* 2012, **31**:2261–2274.
14. Ohno M, Sametsky EA, Younkin LH, Oakley H, Younkin SG, Citron M, Vassar R, Disterhoft JF: **BACE1 deficiency rescues memory deficits and cholinergic dysfunction in a mouse model of Alzheimer's disease**. *Neuron* 2004, **41**:27–33.
15. Ohno M, Chang L, Tseng W, Oakley H, Citron M, Klein WL, Vassar R, Disterhoft JF: **Temporal memory deficits in Alzheimer's mouse models: rescue by genetic deletion of BACE1**. *Eur J Neurosci* 2006, **23**:251–260.
16. Ohno M, Cole SL, Yasvoina M, Zhao J, Citron M, Berry R, Disterhoft JF, Vassar R: **BACE1 gene deletion prevents neuron loss and memory deficits in 5XFAD APP/PS1 transgenic mice**. *Neurobiol Dis* 2007, **26**:134–145.
17. McConlogue L, Buttini M, Anderson JP, Brigham EF, Chen KS, Freedman SB, Games D, Johnson-Wood K, Lee M, Zeller M, Liu W, Motter R, Sinha S: **Partial reduction of BACE1 has dramatic effects on Alzheimer plaque and synaptic pathology in APP transgenic mice**. *J Biol Chem* 2007, **282**:26326–26334.
18. Devi L, Ohno M: **Genetic reductions of β -site amyloid precursor protein-cleaving enzyme 1 and amyloid- β ameliorate impairment of conditioned taste aversion memory in 5XFAD Alzheimer's disease model mice**. *Eur J Neurosci* 2010, **31**:110–118.
19. Kimura R, Devi L, Ohno M: **Partial reduction of BACE1 improves synaptic plasticity, recent and remote memories in Alzheimer's disease transgenic mice**. *J Neurochem* 2010, **113**:248–261.
20. Devi L, Ohno M: **Mechanisms that lessen benefits of β -secretase reduction in a mouse model of Alzheimer's disease**. *Transl Psychiatry* 2013, **3**:e284.
21. Yu H, Saura CA, Choi SY, Sun LD, Yang X, Handler M, Kawarabayashi T, Younkin L, Fedele B, Wilson MA, Younkin S, Kandel ER, Kirkwood A, Shen J: **APP processing and synaptic plasticity in presenilin-1 conditional knock-out mice**. *Neuron* 2001, **31**:713–726.
22. Beglopoulos V, Sun X, Saura CA, Lemere CA, Kim RD, Shen J: **Reduced β -amyloid production and increased inflammatory responses in presenilin deficient knock-out mice**. *J Biol Chem* 2004, **279**:46907–46914.
23. Saura CA, Choi SY, Beglopoulos V, Malkani S, Zhang D, Shankaranarayana Rao BS, Chattarji S, Kelleher RJ, Kandel ER, Duff K, Kirkwood A, Shen J: **Loss of presenilin function causes impairments of memory and synaptic plasticity followed by age-dependent neurodegeneration**. *Neuron* 2004, **42**:23–36.
24. Tabuchi K, Chen G, Sudhof TC, Shen J: **Conditional forebrain inactivation of nicastrin causes progressive memory impairment and age-related neurodegeneration**. *J Neurosci* 2009, **29**:7290–7301.
25. Dewachter I, Reversé D, Caluwaerts N, Ris L, Kuiperi C, Van den Haute C, Spittaels K, Umans L, Serneels L, Thiry E, Moechars D, Mercken M, Godaux E, Van Leuven F: **Neuronal deficiency of presenilin 1 inhibits amyloid plaque formation and corrects hippocampal long-term potentiation but not a cognitive defect of amyloid precursor protein [V717I] transgenic mice**. *J Neurosci* 2002, **22**:3445–3453.
26. Saura CA, Chen G, Malkani S, Choi SY, Takahashi RH, Zhang D, Gouras GK, Kirkwood A, Morris RG, Shen J: **Conditional inactivation of presenilin 1 prevents amyloid accumulation and temporarily rescues contextual and spatial working memory impairments in amyloid precursor protein transgenic mice**. *J Neurosci* 2005, **25**:6755–6764.
27. Melnikova T, Fromholt S, Kim H, Lee D, Xu G, Price A, Moore BD, Golde TE, Felsenstein KM, Savonenko A, Borchelt DR: **Reversible pathologic and cognitive phenotypes in an inducible model of Alzheimer-amyloidosis**. *J Neurosci* 2013, **33**:3765–3779.
28. Sun B, Zhou Y, Halabisky B, Lo I, Cho SH, Mueller-Stieber S, Devidze N, Wang X, Grubb A, Gan L: **Cystatin C-cathepsin B axis regulates amyloid beta levels and associated neuronal deficits in an animal model of Alzheimer's disease**. *Neuron* 2008, **60**:247–257.
29. Fukumoto H, Takahashi H, Tarui N, Matsui J, Tomita T, Hirode M, Sagayama M, Maeda R, Kawamoto M, Hirai K, Terauchi J, Sakura Y, Kakahana M, Kato K, Iwatsubo T, Miyamoto M: **A noncompetitive BACE1 inhibitor TAK-070 ameliorates A β pathology and behavioural deficits in a mouse model of Alzheimer's disease**. *J Neurosci* 2010, **30**:11157–11166.
30. Zhu Z, Li C, Wang X, Yang Z, Chen J, Hu L, Jiang H, Shen X: **2,2',4'-trihydroxychalcone from *Glycyrrhiza glabra* as a new specific BACE1 inhibitor efficiently ameliorates memory impairment in mice**. *J Neurochem* 2010, **114**:374–385.
31. Chang WP, Huang X, Downs D, Cirrito JR, Koelsch G, Holtzman DM, Ghosh AK, Tang J: **β -Secretase inhibitor GRL-8234 rescues age-related cognitive decline in APP transgenic mice**. *FASEB J* 2011, **25**:775–784.
32. Comery TA, Martone RL, Aschmies S, Atchison KP, Diamantidis G, Gong X, Zhou H, Krefl AF, Pangalos MN, Sonnenberg-Reines J, Jacobsen JS, Marquis KL: **Acute γ -secretase inhibition improves contextual fear conditioning in the Tg2476 mouse model of Alzheimer's disease**. *J Neurosci* 2005, **25**:8898–8902.
33. Martone RL, Zhou H, Atchison K, Comery T, Xu JZ, Huang X, Gong X, Jin M, Krefl A, Harrison B, Mayer SC, Aschmies S, Gonzales C, Zaleska MM, Riddell

- DR, Wagner E, Lu P, Sun SC, Sonnenberg-Reines J, Oganessian A, Adkins K, Leach MW, Clarke DW, Huryn D, Abou-Gharbia M, Magolda R, Bard J, Frick G, Raje S, Forlow SB, et al: **Begacestat (GSI-953): a novel selective thiophene sulfonamide inhibitor of amyloid precursor protein γ -secretase for the treatment of Alzheimer's disease.** *J Pharmacol Exp Ther* 2009, **331**:598–608.
34. Netzer WJ, Powell C, Nong Y, Blundell J, Wong L, Duff K, Flajolet M, Greengard P: **Lowering β -amyloid levels rescues learning and memory in a Down syndrome mouse model.** *PLoS one* 2010, **5**:e10943.
35. Mitani Y, Yarimizu J, Saita K, Uchino H, Akashiba H, Shitaka Y, Ni K, Matsuoka N: **Differential effects between γ -secretase inhibitors and modulators on cognitive function in amyloid precursor protein-transgenic and non-transgenic mice.** *J Neurosci*, **32**:2037–2050.
36. Mitani Y, Yarimizu J, Akashiba H, Shitaka Y, Ni K, Matsuoka N: **Amerioration of cognitive deficits in plaque-bearing Alzheimer's disease model mice through selective reduction of nascent soluble A β 42 without affecting other A β pools.** *J Neurochem* 2013, **125**:465–472.
37. Iwatsubo T, Odaka A, Suzuki N, Mizusawa H, Nukina N, Ihara Y: **Visualization of A β 42(43) and A β 40 in senile plaques with end-specific A β monoclonals: evidence that an initially deposited species is A β 42(43).** *Neuron* 1994, **13**:45–53.
38. Cleary JP, Walsh DM, Hofmeister JJ, Shankar GM, Kuskowski MA, Selkoe DJ, Ashe KH: **Natural oligomers of the amyloid-beta protein specifically disrupt cognitive function.** *Nat Neurosci* 2005, **8**:79–84.
39. Lesné S, Koh MT, Kotilinek L, Kaye R, Glabe CG, Yang A, Gallagher M, Ashe KH: **A specific amyloid- β protein assembly in the brain impairs memory.** *Nature* 2006, **440**:352–357.
40. Hasegawa K, Yamaguchi I, Omata S, Gejyo F, Naiki H: **Interaction between A β (1–42) and A β (1–40) in Alzheimer's β -amyloid formation *in vitro*.** *Biochemistry* 1999, **38**:15514–15521.
41. Yan Y, Wang C: **A β 40 protects non-toxic A β 42 monomer from aggregation.** *J Mol Biol* 2007, **369**:909–916.
42. Murray MM, Bernstein SL, Nyugen V, Condron MM, Teplow DB, Bowers MT: **Amyloid β protein: A β 40 inhibits A β 42 oligomerization.** *J Am Chem Soc* 2009, **131**:6316–6317.
43. Watanabe H, Bernier F, Miyagawa T: **A Therapeutic Agent for A β Related Disorders.** Patent publication WO2006/112552.
44. Kuperstein I, Broersen K, Benilova I, Rozenski J, Jonckheere W, Debulpaep M, Vandersteen A, Segers-Nolten I, Van Der Werf K, Subramaniam V, Braeken D, Callewaert G, Bartic C, D'Hooge R, Martins IC, Rousseau F, Schymkowitz J, De Strooper B: **Neurotoxicity of Alzheimer's disease A β peptides is induced by small changes in the A β 42 to A β 40 ratio.** *EMBO J* 2010, **29**:3408–3420.
45. Deng Y, Tarassishin L, Kallhoff V, Peethumongsin E, Wu L, Li YM, Zheng H: **Deletion of presenilin 1 hydrophilic loop sequence leads to impaired γ -secretase activity and exacerbated amyloid pathology.** *J Neurosci* 2006, **26**:3845–3854.
46. Kim J, Onstead L, Randle S, Price R, Smithson L, Zwizinski C, Dickson DW, Golde T, McGowan E: **A β 40 inhibits amyloid deposition *in vivo*.** *J Neurosci* 2007, **27**:627–633.
47. Kim J, Chakrabarty P, Hanna A, March A, Dickson DW, Borchelt DR, Golde T, Janus C: **Normal cognition in transgenic BR12-A β mice.** *Mol Neurodegen* 2013, **8**:15.
48. Tate B, McKee TD, Loureiro RM, Dumin JA, Xia W, Pojasek K, Austin WF, Fuller NO, Hubbs JL, Shen R, Jonker J, Ives J, Bronk B: **Modulation of gamma-secretase for the treatment of Alzheimer's disease.** *Int J Alzheimer Dis* 2012, **2012**:210756.
49. Rogers K, Felsenstein KM, Hrdlicka L, Tu Z, Albayya F, Lee W, Hopp S, Miller MJ, Spaulding D, Yang X, Hodgdon H, Nolan S, Wen M, Costa D, Blain JF, Freeman E, De Strooper B, Vulsteke V, Scrocchi L, Zetterberg H, Portelius E, Hutter-Paier B, Havas D, Ahljianian M, Flood D, Leventhal L, Shapiro G, Patzke H, Chesworth R, Koenig G: **Modulation of γ -secretase by EVP-0015962 reduces amyloid deposition and behavioural deficits in Tg2576 mice.** *Mol Neurodegen* 2012, **7**:61.
50. Imbimbo BP, Del Giudice E, Colavito D, D'Arrigo A, Dalle Carbonare M, Villetti G, Facchinetti F, Volta R, Pietrini V, Baroc MF, Serneels L, De Strooper B, Leon A: **1-(3',4'-dichloro-2-fluoro[1,1'-biphenyl]-4-yl)-cyclopropanecarboxylic acid (CHF5074), a novel γ -secretase modulator, reduces brain β -amyloid pathology in a transgenic mouse model of Alzheimer's disease without causing peripheral toxicity.** *J Pharmacol Exp Ther* 2007, **323**:822–830.
51. Imbimbo BP, Del Giudice E, Cenacchi V, Volta R, Villetti G, Facchinetti F, Riccardi B, Puccini P, Moretto N, Grassi F, Ottonello S, Leon A: ***In vitro* and *in vivo* profiling of CHF5022 and CHF5074, two β -amyloid1-42 lowering agents.** *Pharmacol Res* 2007, **55**:318–328.
52. Imbimbo BP, Hutter-Paier B, Villetti G, Facchinetti F, Cenacchi V, Volta R, Lanzillotta A, Pizzi M, Windisch M: **CHF5074, a novel γ -secretase modulator, attenuates brain β -amyloid pathology and learning deficit in a mouse model of Alzheimer's disease.** *Brit J Pharmacol* 2009, **156**:982–993.
53. Imbimbo BP, Giardino L, Sivilia S, Giuliani A, Gusciglio M, Pietrini V, Del Giudice E, D'Arrigo A, Leon A, Villetti G, Calzà L: **CHF5074, a novel γ -secretase modulator, restores hippocampal neurogenesis potential and reverses contextual memory deficit in a transgenic mouse model of Alzheimer's disease.** *J Alzheimer Dis* 2010, **20**:159–173.
54. Kounnas MZ, Danks AM, Cheng S, Tyree C, Ackerman E, Zhang X, Ahn K, Nguyen P, Comer D, Mao L, Yu C, Pleyne D, Digregorio PJ, Velicelebi G, Stauderman KA, Comer WT, Mobley WC, Li YM, Sisodia SS, Tanzi RE, Wagner SL: **Modulation of γ -secretase reduces β -amyloid deposition in a transgenic mouse model of Alzheimer's disease.** *Neuron* 2010, **67**:769–780.
55. Van Broeck B, Chen JM, Tréton G, Desmidt M, Hopf C, Ramsden N, Karran E, Mercken M, Rowley A: **Chronic treatment with a novel γ -secretase modulator, JN140418677, inhibits amyloid plaque formation in a mouse model of Alzheimer's disease.** *Br J Pharmacol* 2011, **163**:375–389.
56. Wisniewski KE, Boutajangout A: **Immunotherapeutic approaches for Alzheimer's disease in transgenic mouse models.** *Brain Struct Funct* 2010, **214**:201–218.
57. Aisen PS, Vellas B, Harald H: **Moving towards early clinical trials for amyloid-targeted therapy in Alzheimer's disease.** *Nat Rev Drug Disc* 2013, **12**:324.
58. Jack CR Jr, Lowe VJ, Weigand SD, Wiste HJ, Senjem ML, Knopman DS, Shiung MM, Gunter JL, Boeve BF, Kemp BJ, Weiner M, Petersen RC, Alzheimer's Disease Neuroimaging Initiative: **Serial PIB and MRI in normal, mild cognitive impairment and Alzheimer's disease: implications for sequence of pathological events in Alzheimer's disease.** *Brain* 2009, **132**:1355–1365.
59. Jack CR Jr, Wiste HJ, Vemuri P, Weigand SD, Senjem ML, Zeng G, Bernstein MA, Gunter JL, Pankratz VS, Aisen PS, Weiner MW, Petersen RC, Shaw LM, Trojanowski JQ, Knopman DS, Alzheimer's Disease Neuroimaging Initiative: **Brain β -amyloid measures and magnetic resonance imaging atrophy both predict time-to-progression from mild cognitive impairment to Alzheimer's disease.** *Brain* 2010, **133**:3336–3348.
60. Jack CR Jr, Knopman DS, Jagust WJ, Shaw LM, Aisen PS, Weiner MW, Petersen RC, Trojanowski JQ: **Hypothetical model of dynamic biomarkers of the Alzheimer's pathological cascade.** *Lancet Neurol* 2010, **9**:119–128.
61. Jack CR Jr, Vemuri P, Wiste HJ, Weigand SD, Aisen PS, Trojanowski JQ, Shaw LM, Bernstein MA, Petersen RC, Weiner MW, Knopman DS, Alzheimer's Disease Neuroimaging Initiative: **Evidence for ordering of Alzheimer disease biomarkers.** *Arch Neurol* 2011, **68**:1526–1535.
62. Jack CR Jr, Vemuri P, Wiste HJ, Weigand SD, Lesnick TG, Lowe V, Kantarci K, Bernstein MA, Senjem ML, Gunter JL, Boeve BF, Trojanowski JQ, Shaw LM, Aisen PS, Weiner MW, Petersen RC, Knopman DS, Alzheimer's Disease Neuroimaging Initiative: **Shapes of the trajectories of 5 major biomarkers of Alzheimer disease.** *Arch Neurol* 2012, **69**:856–867.
63. Borchhav P, Minthon L, Zetterberg H, Wallin AK, Blennow K, Hansson O: **Cerebrospinal fluid levels of β -amyloid 1–42, but not of tau, are fully changed already 5 to 10 years before the onset of Alzheimer dementia.** *Arch Gen Psychiatry* 2012, **69**:98–106.
64. Bateman RJ, Xiong C, Benzinger TL, Fagan AM, Goate A, Fox NC, Marcus DS, Cairns NJ, Xie X, Blazey TM, Holtzman DM, Santacruz A, Buckles V, Oliver A, Moulder K, Aisen PS, Ghetti B, Klunk WE, McDade E, Martins RN, Masters CL, Mayeux R, Ringman JM, Rossor MN, Schofield PR, Sperling RA, Salloway S, Morris JC, Dominantly Inherited Alzheimer Network: **Clinical and biomarker changes in dominantly inherited Alzheimer's disease.** *N Engl J Med* 2012, **367**:795–804.
65. Mullard A: **Sting of Alzheimer's failures offset by upcoming prevention trials.** *Nat Rev Drug Disc* 2012, **11**:657–660.
66. Carrillo MC, Brashear HR, Logovinsky V, Ryan JM, Feldman HH, Siemers ER, Abushakra S, Hartley DM, Petersen RC, Khachaturian AS, Sperling RA: **Can we prevent Alzheimer's disease? Secondary 'prevention' trials in Alzheimer's disease.** *Alzheimers Dement* 2013, **9**:123–131.
67. Mattsson N, Portelius E, Rolstad S, Gustavsson M, Andreasson U, Stridsberg M, Wallin A, Blennow K, Zetterberg H: **Longitudinal cerebrospinal fluid**

- biomarkers over four years in mild cognitive impairment. *J Alzheimer Dis* 2012, **30**:767–778.
68. Prestia A, Caroli A, van der Flier WM, Ossenkoppele R, Van Berckel B, Barkhof F, Teunissen CE, Wall AE, Carter SF, Schöll M, Choo IH, Nordberg A, Scheltens P, Frisoni GB: **Prediction of dementia in MCI patients based on core diagnostic markers for Alzheimer disease.** *Neurology* 2013, **80**:1048–1056.
69. Benilova I, Karran E, De Strooper B: **The toxic A β oligomer and Alzheimer's disease: an emperor in need of clothes.** *Nat Neurosci* 2012, **15**:349–357.
70. Lu Y, Riddell D, Hajos-Korcsok E, Bales K, Wood KM, Nolan CE, Robshaw AE, Zhang L, Leung L, Becker SL, Tseng E, Barricklow J, Miller EH, Osgood S, O'Neill BT, Brodney MA, Johnson DS, Pettersson M: **CSF A β as an effect biomarker for brain A β lowering verified by quantitative preclinical analyses.** *J Pharmacol Exp Ther* 2012, **342**:366–375.
71. Blennow K, Zetterberg H, Fagan AM: **Fluid biomarkers in Alzheimer disease.** *Cold Spring Harb Perspectives Med* 2012, **2**:a006221.
72. Moreth J, Mavougou C, Schindowski K: **Is A β a sufficient biomarker for monitoring anti-A β clinical studies? A critical review.** *Front Aging Neurosci* 2013, **5**:25.
73. Sutphen CL, Fagan AM, Holtzman DM: **Progress update: fluid and imaging biomarkers in Alzheimer's disease.** *Biol Psychiatry* 2013. doi.org/10.1016/j.biopsych.2013.07.031.
74. Blennow K, Zetterberg H, Rinne JO, Salloway S, Wei J, Black R, Grundman M, Liu E, AAB-001 2011/202 Investigators: **Effect of immunotherapy with bapineuzumab on cerebrospinal fluid biomarker levels in patients with mild to moderate Alzheimer disease.** *Arch Neurol* 2012, **69**:1002–1010.
75. Blennow K, Hampel H, Zetterberg H: **Biomarkers in amyloid- β immunotherapy trials in Alzheimer's disease.** *Neuropsychopharmacology Rev* 2013. 10.1038/npp.2013.154.
76. Lu Y, Zhang L, Nolan CE, Becker SL, Atchison K, Robshaw AE, Pustilnik LR, Osgood SM, Miller EH, Stepan AF, Subramanyam C, Efremov I, Hallgren AJ, Riddell D: **Quantitative pharmacokinetic/pharmacodynamic analyses suggest that 129/SVE mouse is a suitable preclinical pharmacology model for identifying small molecule gamma secretase inhibitors.** *J Pharmacol Exp Ther* 2011, **339**:922–934.
77. Tai LM, Jacobsen H, Ozmen L, Florh A, Jakob-Roetne R, Caruso A, Grimm HP: **The dynamics of A β distribution after γ -secretase inhibitor treatment, as determined by experimental and modeling approaches in a wild type rat.** *J Pharmacokinetic Pharmacodyn* 2012, **229**:227–237.
78. Albright CF, Dockens RC, Meredith JE Jr, Olson RE, Slemmon R, Lentz KA, Wang JS, Denton RR, Pilcher G, Rhyne PW, Raybon JJ, Barten DM, Burton C, Toyn JH, Sankaranarayanan S, Polson C, Guss V, White R, Simutis F, Sanderson T, Gillman KW, Starrett JE Jr, Bronson J, Sverdlow O, Huang SP, Castaneda L, Feldman H, Coric V, Zaczek R, Macor JE, et al: **Pharmacodynamics of selective inhibition of γ -secretase by avagacestat.** *J Pharmacol Exp Ther* 2013, **344**:686–695.
79. Borghys H, Tuefferd M, Van Broeck B, Clessens E, Dillen L, Cools W, Vinken P, Straetmans R, De Ridder F, Gijzen H, Mercken M: **A canine model to evaluate efficacy and safety of γ -secretase inhibitors and modulators.** *J Alzheimer Dis* 2012, **28**:809–822.
80. Hawkins J, Harrison DC, Ahmed S, Davis RP, Chapman T, Marshall I, Smith B, Mead TL, Medhurst A, Giblin GM, Hall A, Gonzalez MI, Richardson J, Hussain I: **Dynamics of A β 42 reduction in plasma, CSF and brain of rats treated with the γ -secretase modulator, GSM-10 h.** *Neurodegen Dis* 2011, **8**:455–464.
81. Best JD, Jay MT, Otu F, Churcher I, Reilly M, Morentin-Gutierrez P, Pattison C, Harrison T, Shearman MS, Atack JR: **In vivo characterization of A β (40) changes in brain and cerebrospinal fluid using the novel β -secretase inhibitor N-[cis-4-[(4-chlorophenyl)sulfonyl]-4-(2,5-difluorophenyl)cyclohexyl]-1,1,1-trifluoromethanesulfonamide (MRK-560) in the rat.** *J Pharmacol Exp Ther* 2006, **317**:786–790.
82. May PC, Dean RA, Lowe SL, Martenyi F, Sheehan SM, Boggs LN, Monk SA, Mathes BM, Mergott DJ, Watson BM, Stout SL, Timm DE, Smith Labell E, Gonzales CR, Nakano M, Jhee SS, Yen M, Ereshefsky L, Lindstrom TD, Calligaro DO, Cocke PJ, Greg Hall D, Friedrich S, Citron M, Audia JE: **Robust central reduction of amyloid- β in humans with an orally available, non-peptidic β -secretase inhibitor.** *J Neurosci* 2011, **31**:16507–16516.
83. Liu X, Wong H, Searce-Lavie K, Watts RJ, Coraggio M, Shin YG, Peng K, Wildsmith KR, Atwal JK, Mango J, Schauer SP, Regal K, Hunt KW, Thomas AA, Siu M, Lyssikatos J, Deshmukh G, Hop CE: **Mechanistic pharmacokinetic-pharmacodynamic modeling of BACE1 inhibition in monkeys: development of a predictive model for amyloid precursor protein processing.** *Drug Metab Disposition Clin Drug Invest* 2013, **41**:1319–1328.
84. Tong G, Castaneda L, Wang JS, Sverdlow O, Huang SP, Slemmon R, Gu H, Wong O, Li H, Berman RM, Smith C, Albright C, Dockens RC: **Effects of single doses of avagacestat (BMS-708163) on cerebrospinal fluid A β levels in healthy young men.** *Clin Drug Invest* 2012, **32**:761–769.
85. Dockens R, Wang JS, Castaneda L, Sverdlow O, Huang SP, Slemmon R, Gu H, Wong O, Li H, Berman RM, Smith C, Albright CF, Tong G: **Placebo-controlled, multiple ascending dose study to evaluate the safety, pharmacokinetics and pharmacodynamics of avagacestat (BMS-708163) in healthy young and elderly subjects.** *Clin Pharmacokinet* 2012, **51**:681–693.
86. Coric V, van Dyck CH, Salloway S, Andreasen N, Brody M, Richter RW, Soininen H, Thein S, Shiovitz T, Pilcher G, Colby S, Rollin L, Dockens R, Pachai C, Portelius E, Andreasson U, Blennow K, Soares H, Albright C, Feldman HH, Berman RM: **Safety and tolerability of the γ -secretase inhibitor avagacestat in a phase 2 study of mild to moderate Alzheimer disease.** *Arch Neurol* 2012, **69**:1430–1440.
87. Wolfe KF, Cyr DM: **Amyloid in neurodegenerative diseases: friend or foe?** *Sem Cell Dev Biol* 2011, **22**:476–481.
88. Spires TL, Meyer-Luehmann M, Stern EA, McLean PJ, Koch J, Nguyen PT, Bacskai BJ, Hyman BT: **Dendritic spine abnormalities in amyloid precursor protein transgenic mice demonstrated by gene transfer and intravital multiphoton microscopy.** *J Neurosci* 2005, **31**:7278–7287.
89. Meyer-Luehmann M, Spires-Jones TL, Prada C, Garcia-Alloza M, de Calignon A, Rozkalne A, Koenigsnecht-Talboo J, Holtzman DM, Bacskai BJ, Hyman BT: **Rapid appearance and local toxicity of amyloid- β plaques in a mouse model of Alzheimer's disease.** *Nature* 2008, **451**:720–724.
90. Xie H, Guan J, Borrelli LA, Xu J, Serrano-Pozo A, Bacskai BJ: **Mitochondrial alterations near amyloid plaques in an Alzheimer's disease mouse model.** *J Neurosci* 2013, **43**:17042–17051.
91. Sperling RA, Laviolette PS, O'Keefe K, O'Brien J, Rentz DM, Pihlajamaki M, Marshall G, Hyman BT, Selkoe DJ, Hedden T, Buckner RL, Becker JA, Johnson KA: **Amyloid deposition is associated with impaired default network function in older persons without dementia.** *Neuron* 2009, **63**:178–188.
92. Sheline YI, Raichle ME, Snyder AZ, Morris JC, Head D, Wang S, Mintun MA: **Amyloid plaques disrupt resting state default mode network connectivity in cognitively normal elderly.** *Biol Psychiatry* 2010, **67**:584–587.
93. Nicoll JA, Barton E, Boche D, Neal JW, Ferrer I, Thompson P, Vlachouli C, Wilkinson D, Bayer A, Games D, Seubert P, Schenk D, Holmes C: **A β species removal after A β 42 immunization.** *J Neuropath Exp Neurol* 2006, **11**:1040–1048.
94. Holmes C, Boche D, Wilkinson D, Yadegarfar G, Hopkins V, Bayer A, Jones RW, Bullock R, Love S, Neal JW, Zotova E, Nicoll JA: **Long-term effects of A β 42 immunisation in Alzheimer's disease: follow-up of a randomised, placebo-controlled phase I trial.** *Lancet* 2008, **372**:216–223.
95. Hong S, Quintero-Monzon O, Ostaszewski BL, Podlisny DR, Cavanaugh WT, Yang T, Holtzman DM, Cirrito JR, Selkoe DJ: **Dynamic analysis of amyloid β -protein in behaving mice reveals opposing changes in ISF versus parenchymal A β during age-related plaque formation.** *J Neurosci* 2011, **31**:15861–15869.
96. Seubert P, Barbour R, Khan K, Motter R, Tang P, Kholodenko D, Kling K, Schenk D, Johnson-Wood K, Schroeter S, Gill D, Jacobsen JS, Pangalos M, Basu G, Games D: **Antibody capture of soluble A β does not reduce cortical A β amyloidosis in the PDAPP mouse.** *Neurodegenerative Dis* 2008, **5**:65–71.
97. Walker JR, Pacoma R, Watson J, Ou W, Alves J, Mason DE, Peters EC, Urbina HD, Welzel G, Althage A, Liu B, Tunland T, Jacobson LH, Harris JL, Schumacher AM: **Enhanced proteolytic clearance of plasma A β by peripherally administered neprilysin does not result in reduced levels of brain A β in mice.** *J Neurosci* 2013, **6**:2457–2464.
98. Stone J, Kleijn HJ, Dockendorf M, Ma L, Palczka J, Tseng J, Tanen M, Forman M: **Consistency of BACE inhibitor-mediated brain amyloid production inhibition by MK-8931 in Alzheimer's patients and healthy young adults.** *Alzheimers Dementia* 2013, **9**(Suppl):P690–P691.
99. Forman MS, Kleijn HJ, Dockendorf M, Palczka J, Tseng J, Canales C, Egan M, Kennedy M, Laterza O, Ma L, Scott J, Tanen M, Apter J, Backonja M, Ereshefsky L, Gevorkyan H, Jhee S, Rynders R, Zari A, Bryan E, Wagner J, Troyer M, Stone J: **The novel BACE inhibitor MK-8931 dramatically lowers CSF beta-amyloid in patients with mild-to-moderate Alzheimer's disease.** *Alzheimers Dementia* 2013, **9**(Suppl):139.

100. Rinne JO, Brooks DJ, Rossor MN, Fox NC, Bullock R, Klunk WE, Mathis CA, Blennow K, Barakos J, Okello AA, Rodriguez Martinez De Liano S, Liu E, Koller M, Gregg KM, Schenk D, Black R, Grundman M: **11C-PIB PET assessment of change in fibrillar amyloid- β load in patients with Alzheimer's disease treated with bapineuzumab: a phase 2, double-blind, placebo-controlled, ascending-dose study.** *Lancet Neurol* 2010, **9**:363–372.
101. Ostrowitzki S, Deptula D, Thurfjell L, Barkhof F, Bohrmann B, Brooks DJ, Klunk WE, Ashford E, Yoo K, Xu ZX, Loetscher H, Santarelli L: **Mechanism of amyloid removal in patients with Alzheimer disease treated with gantenerumab.** *Arch Neurol* 2012, **69**:198–207.
102. Siemers ER, Dean RA, Friedrich S, Ferguson-Sells L, Gonzales C, Farlow MR, May PC: **Safety, tolerability and effects on plasma and cerebrospinal fluid amyloid-beta after inhibition of γ -secretase.** *Clin Neuropharmacol* 2007, **30**:317–325.
103. Siemers ER, Quinn JF, Kaye J, Farlow MR, Porsteinsson A, Tariot P, Zoulnouni P, Galvin JE, Holtzman DM, Knopman DS, Sattenwhite J, Gonzales C, Dean RA, May PC: **Effects of a γ -secretase inhibitor in a randomized study of patients with Alzheimer disease.** *Neurology* 2006, **66**:602–604.
104. Fleisher AS, Raman R, Siemers ER, Becerra L, Clark CM, Dean RA, Farlow MR, Galvin JE, Peskind ER, Quinn JF, Sherzai A, Sowell BB, Aisen PS, Thal LJ: **Phase 2 safety trial targeting amyloid β production with a γ -secretase inhibitor in Alzheimer disease.** *Arch Neurol* 2008, **65**:1031–1038.
105. Bateman RJ, Siemers ER, Mawuenyega KG, Wen G, Browning KR, Sigurdson WC, Yarasheski KE, Friedrich SW, Demattos RB, May PC, Paul SM, Holtzman DM: **A β -secretase inhibitor decreases amyloid- β production in the central nervous system.** *Ann Neurol* 2009, **66**:48–54.
106. Portelius E, Dean RA, Gustavsson MK, Andreasson U, Zetterberg H, Siemers E, Blennow K: **A novel A β isoform pattern in CSF reflects γ -secretase inhibition in Alzheimer disease.** *Alzheimer Res Ther* 2010, **2**:7.
107. Portelius E, Zetterberg H, Dean RA, Marcil A, Bourgeois P, Nutu M, Andreasson U, Siemers E, Mawuenyega KG, Sigurdson WC, May PC, Paul SM, Holtzman DM, Blennow K, Bateman RJ: **Amyloid- β 1-15/16 as a marker for γ -secretase inhibition in Alzheimer's disease.** *J Alzheimer Dis* 2012, **31**:335–341.
108. Doody RS, Raman R, Farlow M, Iwatsubo T, Vellas B, Joffe S, Kiebertz K, He F, Sun X, Thomas RG, Aisen PS, Alzheimer's Disease Cooperative Study Steering Committee, Siemers E, Sethuraman G, Mohs R, Semagacestat Study Group: **A phase 3 trial of semagacestat for treatment of Alzheimer's disease.** *N Engl J Med* 2013, **369**:341–350.
109. Green RC, Schneider LS, Amato DA, Beelen AP, Wilcock G, Swabb EA, Zavitz KH, Tarenfluril Phase 3 Study Group: **Effect of tarenfluril on cognitive decline and activities of daily living in patients with mild Alzheimer disease: a randomized controlled trial.** *JAMA* 2009, **302**:2557–2564.
110. Eriksen JL, Sagi SA, Smith TE, Weggen S, Das P, McLendon DC, Ozols VV, Jessing KW, Zavitz KH, Koo EH, Golde TE: **NSAIDs and enantiomers of flurbiprofen target γ -secretase and lower A β 42 in vivo.** *J Clin Invest* 2003, **112**:440–449.
111. Lanz TA, Fici GJ, Merchant KM: **Lack of specific amyloid- β (1–42) suppression by nonsteroidal anti-inflammatory drugs in young, plaque-free Tg2576 mice and in guinea pig neuronal cultures.** *J Pharmacol Exp Ther* 2005, **312**:399–406.
112. Ross J, Sharma S, Winston J, Nunez M, Bottini G, Franceschi M, Scarpini E, Frigerio E, Fiorentini F, Fernandez M, Sivilia S, Giardino L, Calza L, Norris D, Cicirello H, Casula D, Imbimbo BP: **CHF5074 reduces biomarkers of neuroinflammation in patients with mild cognitive impairment: a 12-week, double-blind, placebo-controlled study.** *Curr Alzheimer Res* 2013, **10**:742–753.
113. Pettersson M, Stepan AF, Kauffman GW, Johnson DS: **Novel γ -secretase modulators for the treatment of Alzheimer's disease: a review focusing on patents from 2010 to 2012.** *Expert Opin Ther Pat* 2013, **23**:1349–1366.
114. Schenk D, Barbour R, Dunn W, Gordon G, Grajeda H, Guido T, Hu K, Huang J, Johnson-Wood K, Khan K, Kholodenko D, Lee M, Liao Z, Lieberburg I, Motter R, Mutter L, Soriano F, Shopp G, Vasquez N, Vandeventer C, Walker S, Wogulis M, Yednock T, Games D, Seubert P: **Immunization with amyloid- β attenuates Alzheimer-disease-like pathology in the PDAPP mice.** *Nature* 1999, **400**:173–177.
115. Gilman S, Koller M, Black RS, Jenkins L, Griffith SG, Fox NC, Eisner L, Kirby L, Rovira MB, Forette F, Orgogozo JM, AN1792(QS-21)-201 Study Team: **Clinical effects of A β immunization (AN1792) in patients with AD in an interrupted trial.** *Neurology* 2005, **64**:1553–1562.
116. Salloway S, Sperling R, Fox NC, Blennow K, Klunk W, Raskind M, Sabbagh M, Honig LS, Porsteinsson AP, Ferris S, Reichert M, Ketter N, Nejadnik B, Guenzler V, Miloslavsky M, Wang D, Lu Y, Lull J, Tudor IC, Liu E, Grundman M, Yuen E, Black R, Brashear HR, Bapineuzumab 301 and 302 Clinical Trial Investigators: **Two phase 3 trials of bapineuzumab in mild-to-moderate Alzheimer's disease.** *N Engl J Med* 2014, **370**:322–333.
117. Doody RS, Thomas RG, Farlow M, Iwatsubo T, Vellas B, Joffe S, Kiebertz K, Raman R, Sun X, Aisen PS, Siemers E, Liu-Seifert H, Mohs R, Alzheimer's Disease Cooperative Study Steering Committee, Solanezumab Study Group: **Phase 3 trials of solanezumab for mild-to-moderate Alzheimer's disease.** *N Engl J Med* 2014, **370**:311–321.
118. Levites Y, Smithson LA, Price RW, Dakin RS, Yuan B, Sierks MR, Kim J, McGowan E, Reed DK, Rosenberry TL, Das P, Golde TE: **Insights into the mechanisms of action of anti-A β antibodies in Alzheimer's disease mouse models.** *FASEB J* 2006, **20**:E2002–E2014.
119. Golde T, Das P, Levites Y: **Quantitative and mechanistic studies of A β immunotherapy.** *CNS Neurol Disord Drug Targets* 2009, **8**:31–49.
120. Demattos RB, Lu J, Tang Y, Racke MM, DeLong CA, Tzaferis JA, Hole JT, Forster BM, McDonnell PC, Liu F, Kinley RD, Jordan WH, Hutton ML: **A plaque-specific antibody clears existing β -amyloid plaques in Alzheimer's disease mice.** *Neuron* 2012, **76**:908–920.
121. Streffer JR, Blennow K, Salloway S, Zetterberg H, Xu Y-Z, Lu Y, Lull J, Collins P, Tudor IC, Gregg K, Styren S, Yuen E, Grundman M, Brashear RH, Liu E: **Effects of bapineuzumab on CSF p-Tau and t-Tau in mild-to-moderate Alzheimer's disease: results from two phase III trials in APO- ϵ 4 carriers and non-carriers.** *Alzheimers Dementia* 2013, **9**(Suppl):138.
122. Farlow M, Arnold SE, van Dyck CH, Aisen PS, Snider BJ, Porsteinsson AP, Friedrich S, Dean RA, Gonzales C, Sethuraman G, DeMattos RB, Mohs R, Paul SM, Siemers ER: **Safety and biomarker effects of solanezumab in patients with Alzheimer's disease.** *Alzheimers Dement* 2012, **8**:261–271.
123. Puli L, Pomeschchik Y, Olas K, Malm T, Koistinaho J, Tanila H: **Effects of human intravenous immunoglobulin on amyloid pathology and neuroinflammation in a mouse model of Alzheimer's disease.** *J Neuroinflamm* 2012, **9**:105.
124. Sudduth TL, Greenstein A, Wilcock DM: **Intracranial injection of gammagard, a human IVIg, modulates the inflammatory response of the brain and lowers A β in APP/PS1 mice along a different time course than anti-A β antibodies.** *J Neurosci* 2013, **33**:9684–9692.
125. Relkin NR, Szabo P, Adamiak B, Burgut T, Monthe C, Lent RW, Younkin S, Younkin L, Schiff R, Weksler ME: **18-Month study of intravenous immunoglobulin for treatment of mild Alzheimer disease.** *Neurobiol Aging* 2009, **30**:1728–1736.
126. Dodel R, Rominger A, Bartenstein P, Barkhof F, Blennow K, Förster S, Winter Y, Bach JP, Popp J, Alferink J, Wiltfang J, Buerger K, Otto M, Antuono P, Jacoby M, Richter R, Stevens J, Melamed I, Goldstein J, Haag S, Wietek S, Farlow M, Jessen F: **Intravenous immunoglobulin for treatment of mild-to-moderate Alzheimer's disease: a phase 2, randomised, double-blind, placebo-controlled, dose-finding trial.** *Lancet Neurol* 2013, **12**:233–243.
127. Adolfsen O, Pihlgren M, Toni N, Varisco Y, Buccarello AL, Antonello K, Lohmann S, Piorkowska K, Gafner V, Atwal JK, Maloney J, Chen M, Gogineni A, Weimer RM, Mortensen DL, Friesenhahn M, Ho C, Paul R, Pfeifer A, Muhs A, Watts RJ: **An effector-reduced anti- β -amyloid (A β) antibody with unique A β binding properties promotes neuroprotection and glial engulfment of A β .** *J Neurosci* 2012, **32**:9677–9689.
128. Bohrmann B, Baumann K, Benz J, Gerber F, Huber W, Knoflach F, Messer J, Oroszlan K, Rauchenberger R, Richter WF, Rothe C, Urban M, Bardroff M, Winter M, Nordstedt C, Loetscher H: **Gantenerumab: a novel human anti-A β antibody demonstrates sustained cerebral amyloid- β binding and elicits cell-mediated removal of human amyloid- β .** *J Alzheimer Dis* 2011, **26**:1–21.

10.1186/alzrt244

Cite this article as: Toyn and Ahljianian: Interpreting Alzheimer's disease clinical trials in light of the effects on amyloid- β . *Alzheimer's Research & Therapy* 2014, **6**:14