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The use of synaptic biomarkers in cerebrospinal fluid to differentiate behavioral variant of frontotemporal dementia from primary psychiatric disorders and Alzheimer's disease

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Abstract

Background Lack of early molecular biomarkers in sporadic behavioral variants of frontotemporal dementia (bvFTD) and its clinical overlap with primary psychiatric disorders (PPD) hampers its diagnostic distinction. Synaptic dysfunction is an early feature in bvFTD and identification of specific biomarkers might improve its diagnostic accuracy. Our goal was to understand the differential diagnostic potential of cerebrospinal fluid (CSF) synaptic biomarkers in bvFTD versus PPD and their specificity towards bvFTD compared with Alzheimer's disease (AD) and controls. Additionally, we explored the association of CSF synaptic biomarkers with social cognition, cognitive performance, and disease severity in these clinical groups.

Methods Participants with probable bvFTD ($n = 57$), PPD ($n = 71$), AD ($n = 60$), and cognitively normal controls ($n = 39$) with available CSF, cognitive tests, and disease severity as frontotemporal lobar degeneration-modified clinical dementia rating scale (FTLD-CDR) were included. In a subset of bvFTD and PPD cases, Ekman 60 faces test scores for social cognition were available. CSF synaptosomal-associated protein 25 (SNAP25), neurogranin (Ng), neuronal pentraxin 2 (NPTX2), and glutamate receptor 4 (GluR4) were measured, along with neurofilament light (NfL), and compared between groups using analysis of covariance (ANCOVA) and logistic regression. Diagnostic accuracy was assessed using ROC analyses, and biomarker panels were selected using Wald's backward selection. Correlations with cognitive measures were performed using Pearson's partial correlation analysis.

Results NPTX2 concentrations were lower in the bvFTD group compared with PPD ($p < 0.001$) and controls ($p = 0.003$) but not compared with AD. Concentrations of SNAP25 ($p < 0.001$) and Ng ($p < 0.001$) were elevated in patients with AD versus those with bvFTD and controls. The modeled panel for differential diagnosis of bvFTD versus PPD consisted of NfL and NPTX2 (AUC = 0.96, CI: 0.93–0.99, $p < 0.001$). In bvFTD versus AD, the modeled panel consisted of NfL, SNAP25, Ng, and GluR4 (AUC = 0.86, CI: 0.79–0.92, $p < 0.001$). In bvFTD, lower NPTX2 (Pearson's $r = 0.29$, $p = 0.036$) and GluR4 (Pearson's $r = 0.34$, $p = 0.014$) concentrations were weakly associated with worse performance

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of total cognitive score. Lower GluR4 concentrations were also associated with worse MMSE scores (Pearson's $r = 0.41$, $p = 0.002$) as well as with worse executive functioning (Pearson's $r = 0.36$, $p = 0.011$) in bvFTD. There were no associations between synaptic markers and social cognition or disease severity in bvFTD.

Conclusion Our findings of involvement of NPTX2 in bvFTD but not PPD contribute towards better understanding of bvFTD disease pathology.

Keywords Synaptic biomarkers, Frontotemporal dementia, Primary psychiatric disorders, Differential diagnosis

Background

Frontotemporal dementia (FTD) is a clinically, genetically, and pathologically heterogeneous disease and the second most common form of young-onset dementia after Alzheimer's disease (AD) [1]. The most prevalent form is the behavioral variant of FTD (bvFTD), which is characterized by slowly progressive behavioral symptoms and impaired social cognition. In 30% of the bvFTD patients, the disease is caused by a pathogenic mutation (C9ORF72, MAPT, GRN), but the majority of cases are denoted sporadic bvFTD (70%) [2]. The lack of an early, disease-specific molecular biomarker in sporadic bvFTD together with its significant overlap in clinical symptoms with primary psychiatric disorders (PPD) and frontotemporal hypometabolic patterns on [^{18}F]-2-deoxy-2-fluoro-D-glucose (FDG) positron emission tomography ([^{18}F]FDG-PET) scans hampers diagnostic distinction. This leads to misdiagnosis in 50% of the cases with an average diagnostic delay of 6.4 years [3–5].

Evidence from post-mortem and clinical studies including pre-dementia stages show that loss of synaptic function is a predominant early feature in bvFTD and correlates with the level of cognitive impairment [6–14]. In accordance, a recent in vivo study using the [^{11}C]UCB-J PET tracer to detect synaptopathy showed widespread frontotemporal loss of synapses in symptomatic bvFTD patients, which is related to disease severity [14]. Also, lower brain synaptic densities on [^{18}F]UCB-H PET were found in the temporal brain regions involved in social cognition, highlighting the clinical relevance of synaptopathy in the disease pathophysiology of FTD [11].

In agreement, using cerebrospinal fluid (CSF) biomarkers as indicators for synapse health, differential synaptic concentrations were found in genetic forms of FTD [12]. Previous studies on synaptic involvement in FTD predominately assessed genetic FTD cases, had a small sample size, or did not include PPD as a control group, whereas the latter is the most challenging to distinguish from bvFTD in clinical practice [10–12]. Identification of specific CSF synaptic markers in sporadic bvFTD might improve diagnostic accuracy and aid in a better understanding of FTD pathophysiology. In addition, specific CSF-synaptic panels can provide endpoints in future

clinical trials of sporadic bvFTD as they might correlate and reflect cognitive and social functioning.

In an attempt to explore the synaptic pathology in FTD and AD, we recently performed a pilot study where we found that concentrations of CSF synaptic biomarkers synaptosomal-associated protein 25 (SNAP25) and neurogranin (Ng) were elevated in FTD compared with controls, while those of neuronal pentraxin 2 (NPTX2) were lower than in controls, suggesting these could be valuable biomarkers for FTD [15]. SNAP25 is a pre-synaptic vesicle protein involved in neurotransmission, while Ng in the post-synapse regulates calcium ion influxes, and NPTX2 present extracellularly in the synaptic cleft maintains synaptic plasticity [15]. Simultaneously, another recent study suggested that patients with primary psychiatric disorders had significantly lower expression of the post-synaptic protein glutamate receptor 4 (GluR4) in CSF compared with cognitively normal controls and therefore could be useful as biomarkers for PDD [16]. GluR4 is primarily involved in excitatory signal transmission [15]. The axonal protein neurofilament light chain (NfL) has emerged as a promising fluid biomarker to distinguish bvFTD from PPD. Several studies have reported the potential of NfL as a biomarker that correlates with brain atrophy, neurodegeneration, and cognition in dementias [17–20] and also with other neuronal damage, e.g., due to stroke, amyotrophic lateral sclerosis, and multiple sclerosis [3, 21–24].

In this study, we aimed to assess the diagnostic performance of CSF synaptic biomarkers in sporadic bvFTD versus PPD and their added value compared with NfL as well as their specificity towards bvFTD compared with AD and controls. Secondly, we assessed the association of CSF synaptic biomarkers with social cognition, cognitive performance, and disease severity in bvFTD.

Methods

Participants

Patients with sporadic probable bvFTD, PPD, AD, and cognitively normal controls who visited the memory clinic of the Alzheimer Center Amsterdam between 2003 and 2021 were included in this study [25–27]. We included individuals aged 45–75 years with available CSF in the biobank and available clinical data. Individuals

with AD were included in the case of AD-positive CSF biomarkers. bvFTD, PPD, and controls were excluded in the case of AD-positive CSF biomarkers. The study was approved by the Medical Ethical Committee of Amsterdam UMC. All participants provided informed consent and the study has been carried out by the Declaration of Helsinki.

Diagnostic procedure

All participants had an extensive, standardized diagnostic assessment including clinical evaluation by a cognitive neurologist and/or old age psychiatrist, blood examination to exclude somatic causes, administered neuropsychological tests assessing five cognitive domains (attention, memory, speed, executive functioning, visuospatial functioning) [28, 29], lumbar puncture for CSF assessment of the AD biomarkers amyloid-beta42, total tau, and phosphorylated tau181 to determine positive or negative AD biomarkers status (cutoffs applied as published elsewhere), electroencephalography and neuroimaging- magnetic resonance imaging, and, if indicated, a [¹⁸F]FDG-PET scan [3, 30]. The diagnosis was concluded in a multidisciplinary meeting using consensus criteria for probable FTD, PPD (DSM-V), and AD [25, 31, 32]. Controls had no evidence of current or recent psychiatric disorders nor a neurodegenerative disorder. Psychiatric diagnoses included mood disorders (*n* =33), personality disorders (*n* = 4), autism spectrum disorder (*n* = 3), anxiety disorder (*n* = 4), functional disorder (*n* = 5), schizophrenia (*n* = 2), and other psychiatry (*n* = 10) [33]. The psychotropic medications the patients were taking included antidepressants, mood stabilizers (lithium), benzodiazepines, amphetamines (methylphenidate), antiepileptic and antipsychotic drugs, and cholinesterase inhibitors (rivastigmine).

Measures for cognition, disease severity, mood and behavioral symptoms

Participants underwent standardized neuropsychological assessments covering the indicated cognitive domains (memory, attention, executive functioning, language,

visuospatial functioning) [29]. For each cognitive domain, at least two tests were used to provide a reliable outcome on cognitive functioning (Table 1) [28]. Per test, a z-score was calculated, and, subsequently, all z-scores covering a specific cognitive domain were averaged into a cognitive domain z-score. Total cognitive score was calculated as an average z-score based on all five cognitive domains (memory, attention, executive functioning, language, visuospatial functioning, Table 1).

A subgroup of the bvFTD group (*n* = 13) and the PPD group (*n* = 9) completed a facial emotional recognition test (the Ekman 60 faces test). Disease severity in bvFTD was scaled according to the Frontotemporal Lobar Degeneration-Modified Clinical Dementia Rating (FTLD-CDR) Scale, using the sum-of-boxes score [34]. Also, participants completed the Geriatric Depression Scale (GDS) (mood), the Frontal Assessment Battery (FAB) (behavioral symptoms), and the Mini-Mental State Exam (MMSE) for global cognition. All tests and FTLD-CDR were completed within 12 months of the CSF withdrawal.

CSF biomarker measurements

NfL was measured using a novel ELISA developed at ADx NeuroSciences, described elsewhere [35]. Synaptic protein Ng was measured using a commercial ELISA from EuroImmun, while the other synaptic proteins SNAP25, NPTX2, and GluR4 were measured using novel immunoassays developed and validated as per standardized protocols at ADx NeuroSciences, described in detail elsewhere [15, 36, 37]. The biomarkers are stable up to at least four freeze-thaw cycles in CSF. Thus, we measured SNAP25 and Ng in the first freeze-thaw cycle of the CSF samples, NfL in the second, NPTX2 in the third, and GluR4 in the final fourth freeze-thaw cycle. All clinical duplicate measurements were well within the range of 20% coefficient of variation (CV) for all immunoassays, except GluR4 which had slightly higher variability. The intermediate precision (average %CV) for each immunoassay of quality control samples was as follows:

Table 1 The cognitive tests included to calculate the z-scores for each cognitive domain [29]

Cognitive domains	Tests included
Memory	Visual association test (VAT), Dutch version of the Rey auditory verbal learning test (RAVLT) with subtests of total immediate recall and delayed recall
Attention	Digit span forward, trail-making test (TMT) A, Stroop color-word test I and II
Executive functioning	Stroop color-word test III, digit span backwards, frontal assessment battery (FAB), letter fluency test (version D-A-T)
Language	Visual association test (VAT)—“naming,” category fluency animals
Visuospatial functioning	Visual object and space perception (VOSP) battery: number location, dot counting, fragmented letters
Total cognitive score	Memory, attention, executive functioning, language, visuospatial functioning

NfL—12%, SNAP25—4%, Ng—11%, NPTX2—15%, and GluR4—25%.

Statistics

All statistical analyses were performed using IBM SPSS Statistics (v.28.0.1.1) or RStudio (v.4.0.3). The demographic differences between the diagnostic groups were assessed using a one-way analysis of variance (ANOVA) or chi-square test where appropriate. The CSF biomarker concentrations and cognitive test scores were log10 transformed to fit a normal distribution. Analysis of covariance (ANCOVA) models corrected for age, sex, and psychotropic medication use with post hoc pairwise comparisons was used to determine differences in biomarker concentrations between the clinical groups, with Bonferroni's multiple comparison correction. Next, logistic regression analysis was performed to assess the association between the CSF biomarkers and diagnosis for the groups bvFTD versus PPD and bvFTD versus AD while controlling for the effect of age, sex, and psychotropic medication use. Additionally, we used Wald's

backward logistic regression on the biomarkers to select a biomarker panel for the group comparisons between bvFTD and AD or PPD. Receiver operating characteristics (ROC) curves were constructed for the CSF synaptic biomarkers and CSF NfL, as well as for the biomarker panels, not controlling for any potential confounders. For each ROC curve, the sensitivity and specificity were determined at Youden's indices. Correlation between the CSF synaptic biomarkers and CSF NfL and cognitive test scores were assessed using Pearson's partial correlation analysis controlling for age. Significance was defined as $p < 0.05$.

Results

Demographic characteristics

The demographic characteristics of the patients are detailed in Table 2. This cohort included 57 patients with bvFTD (age: 64 ± 8 , female: 37%), 71 patients with PPD (age: 56 ± 9 , female: 38%), 60 patients with AD (age: 66 ± 7 , female: 45%), and 39 controls (age: 57 ± 8 , female: 33%). The bvFTD and AD groups were significantly older

Table 2 Demographic characteristics of the study cohort

Diagnostic groups	bvFTD (n = 57)	PPD (n = 71)	AD (n = 60)	Controls (n = 39)	p-value
Age	64 (8) ^{b,d}	56 (9) ^{a,c}	66 (7) ^{b,d}	57 (8) ^{a,c}	< 0.001
Female sex (%)	21 (37%)	27 (38%)	27 (45%)	13 (33%)	0.737
Psychotropic medication use (% Yes)	16 (30%)	36 (51%) ^d	21 (35%)	5 (15%) ^b	0.014
CSF biomarkers (pg/mL)					
NfL	1630 (1052) ^{b,c,d}	369 (178) ^{a,c}	848 (332) ^{a,b,d}	337 (187) ^{a,c}	< 0.001
SNAP25	37 (32) ^c	33 (22) ^c	51 (19) ^{a,b,d}	28 (7) ^c	< 0.001
Ng	389 (312) ^c	351 (230) ^c	723 (1069) ^{a,b,d}	283 (107) ^c	< 0.001
NPTX2	401 (270) ^{b,d}	583 (300) ^a	477 (227) ^b	542 (240) ^a	< 0.001
GluR4	1100 (995)	1223 (1070)	1112 (510)	1070 (386)	0.456
Cognitive domains					
Memory	- 0.13 (0.6) ^{b,c,d}	0.41 (0.6) ^{a,c}	- 0.95 (0.7) ^{a,b,d}	0.73 (0.6) ^{a,c}	< 0.001
Attention	- 0.35 (1.0) ^{b,d}	0.14 (0.8) ^{a,c,d}	- 0.27 (0.8) ^{b,d}	0.53 (0.3) ^{a,b,c}	< 0.001
Executive functioning	- 0.44 (0.9) ^{b,d}	0.16 (0.6) ^{a,c,d}	- 0.43 (0.8) ^{b,d}	0.64 (0.4) ^{a,b,c}	< 0.001
Language	- 0.29 (0.7) ^{b,d}	0.32 (0.5) ^{a,c}	- 0.44 (0.9) ^{b,d}	0.60 (0.5) ^{a,c}	< 0.001
Visuospatial functioning	0.11 (0.5) ^c	0.19 (0.5) ^c	- 0.64 (1.1) ^{a,b,d}	0.34 (0.3) ^c	< 0.001
Total cognitive score	- 0.43 (0.8) ^{b,d}	0.2 (0.7) ^{a,c,d}	- 0.60 (0.7) ^{b,d}	0.60 (0.3) ^{a,b,c}	< 0.001
Other tests					
MMSE	24.3 (5) ^{b,d}	26.9 (2) ^{a,c}	20.7 (5) ^{b,d}	28.3 (1) ^{a,c}	< 0.001
Geriatric depression scale	3.2 (3) ^b	6.3 (4) ^{a,c,d}	2.9 (3) ^b	3.4 (3) ^b	< 0.001
Ekman 60 faces test	33.7 (7) ^b	42.7 (7) ^a	-	-	0.023
FTLD- CDR	7 (4)	-	-	-	-

Cognitive domain data are represented by z-scores. Other tests are represented as absolute scores. Missing data: GluR4: 3 bvFTD, 3 AD, 1 control; memory: 2 AD, 9 bvFTD, 3 PPD; attention: 2 bvFTD; executive functioning: 2 AD, 4 bvFTD, 2 PPD; visuospatial functioning: 22 AD, 25 bvFTD, 11 PPD, 4 controls; total cognitive score: 2 bvFTD; MMSE: 3 AD, 1 bvFTD, 1 PPD; geriatric depression scale: 7 AD, 11 bvFTD, 6 PPD, 2 controls; Ekman 60 faces test: 44 bvFTD, 62 PPD (not available for AD and controls); FTLD-CDR (available for bvFTD only): 2 bvFTD

bvFTD, behavioral frontotemporal dementia; *PPD*, primary psychiatric disorders; *AD*, Alzheimer's disease; *NfL*, neurofilament light; *SNAP25*, synaptosomal-associated protein 25; *Ng*, neurogranin; *NPTX2*, neuronal pentraxin 2; *MMSE*, mini-mental state examination; *FTLD-CDR*, frontotemporal lobar degeneration-modified clinical dementia rating scale

Data represents the mean (SD) or n (%); ^a $p < 0.05$ vs bvFTD, ^b $p < 0.05$ vs PPD, ^c $p < 0.05$ vs AD, ^d $p < 0.05$ vs controls

than the PPD group and controls ($p < 0.001$ for all). No group differences were found for sex. Thirty percent of patients with bvFTD, 51% of those with PPD, 35% of the AD group, and 15% of the controls used psychotropic medication. Following expectations, psychotropic medication use was more frequent in the PPD group ($p < 0.001$) than in the control group.

Among the cognitive domain scores, the bvFTD group scored significantly ($p < 0.05$) lower than those with PPD for all domains except visuospatial functioning. Furthermore, patients with AD had significantly lower memory ($p < 0.001$) and visuospatial functioning ($p = 0.004$) scores than the bvFTD group. As expected, the bvFTD group had lower MMSE scores than PPD ($p = 0.018$) or controls ($p = 0.004$), while the AD group had the lowest MMSE scores, the difference being significant compared with PPD ($p < 0.001$) or controls ($p < 0.001$) but not bvFTD. Following expectations, the Ekman 60 faces test scores were lower in bvFTD patients compared with PPD patients ($p = 0.012$) (not available for AD and controls).

All bvFTD patients in this cohort had mild to moderate FTD disease severity (FTLD-CDR ≤ 16).

Differential concentrations of candidate CSF biomarkers across the diagnostic groups

The concentrations of the CSF biomarkers are presented in Table 2 and visualized in Fig. 1. Adjusted for age, sex, and psychotropic medication use, we found that NPTX2 concentrations were significantly lower in bvFTD compared with PPD ($p < 0.001$) and controls ($p = 0.005$). There was a trend towards lower average NPTX2 concentrations in bvFTD than those in the AD group, although this difference was not significant. SNAP25 and Ng concentrations were higher in AD compared with bvFTD, PPD, and controls ($p < 0.001$ for all). CSF GluR4 concentrations did not differ between the diagnostic groups. Furthermore, NfL concentrations were higher in bvFTD compared with AD, PPD, and controls ($p < 0.001$ for all). NfL did not differ

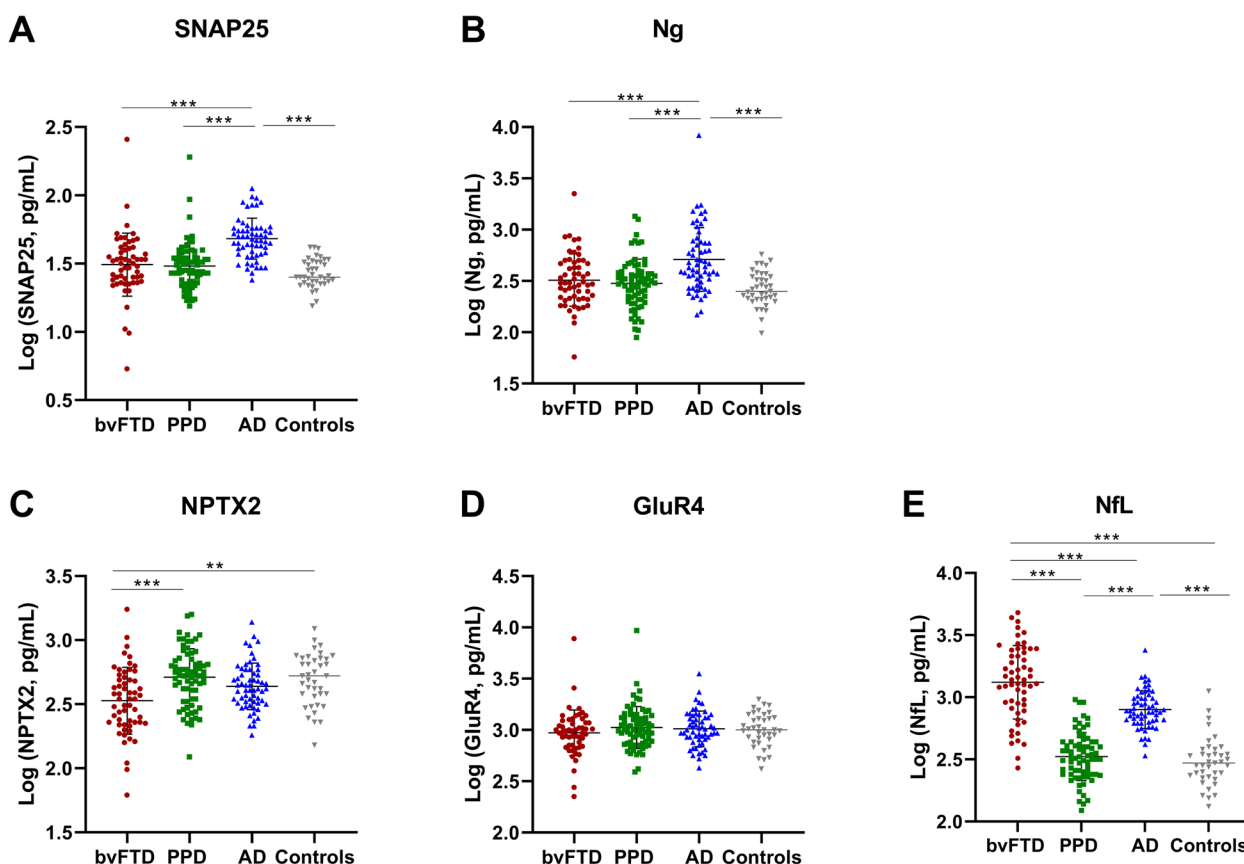


Fig. 1 Concentrations of candidate CSF biomarkers across the diagnostic groups. Analysis of covariance (ANCOVA) models corrected for age, sex, and psychotropic medication use with post hoc pairwise comparisons were used to determine the log10 transformed biomarker differences between the clinical groups, with Bonferroni's multiple comparison correction. **A** SNAP25. **B** Ng. **C** NPTX2. **D** GluR4. **E** NfL. SNAP25, synaptosomal-associated protein 25; Ng, neurogranin; NPTX2, neuronal pentraxin 2; GluR4, glutamate receptor 4; NfL, neurofilament light. bvFTD, behavioral frontotemporal dementia; PPD, primary psychiatric disorders; AD, Alzheimer's disease. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$

between PPD and controls, while patients with AD had higher NfL concentrations compared with PPD and controls ($p < 0.001$ for both).

The logistic regression models for each biomarker controlled for age, sex, and psychotropic medication use, and their predictive value for bvFTD compared with PPD and AD are detailed in Table 3. Of the synaptic biomarkers, only NPTX2 had a significant predictive value for bvFTD versus PPD (odds ratio, OR: 0.997 [0.996, 0.999], $p = 0.007$). The synaptic biomarkers SNAP25 (OR: 0.966 [0.943, 0.990], $p = 0.005$) and Ng (OR: 0.998 [0.997, 0.999], $p = 0.006$) had significant diagnostic values for bvFTD versus AD.

Diagnostic performance of the candidate CSF biomarkers

We further investigated the diagnostic potential of the CSF biomarkers, stand-alone, as well as statistically selected panels, uncorrected for possible confounders (Table 4, Fig. 2).

Table 3 Predictive value of the CSF biomarkers for bvFTD versus PPD and AD

Biomarker	bvFTD vs PPD	bvFTD vs AD
	OR (95% CI)	OR (95% CI)
NfL	1.006 (1.003, 1.009)***	1.002 (1.001, 1.003)***
SNAP25	0.999 (0.986, 1.013)	0.966 (0.943, 0.990)**
Ng	1.000 (0.998, 1.001)	0.998 (0.997, 0.999)**
NPTX2	0.997 (0.996, 0.999)**	0.999 (0.997, 1.000)
GluR4	1.000 (0.999, 1.000)	1.000 (0.999, 1.000)

The logistic regression models for each biomarker are controlled for confounders' age, sex, and psychotropic medication use. OR, odds ratio; NfL, neurofilament light; SNAP25, synaptosomal-associated protein 25; Ng, neurogranin; NPTX2, neuronal pentraxin 2; GluR4, glutamate receptor 4; bvFTD, behavioral frontotemporal dementia; PPD, primary psychiatric disorders; AD, Alzheimer's disease. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$

bvFTD versus PPD

Among the synaptic biomarkers, NPTX2 had the highest AUC (AUC= 0.72, CI: 0.63–0.81, $p < 0.001$) to discriminate bvFTD from PPD. SNAP25, Ng, and GluR4 were not predictive of the diagnostic group. The AUC of NfL (AUC= 0.95, CI: 0.91–0.99, $p < 0.001$) was higher than that of the synaptic proteins. The biomarker panel to differentiate bvFTD from PPD, selected using Wald's backward selection among the candidate biomarkers consisted of NfL and NPTX2 (AUC =0.96, CI: 0.93–0.99, $p < 0.001$).

bvFTD versus AD

Among the synaptic biomarkers, SNAP25 had the highest AUC (AUC = 0.79, CI: 0.70–0.87, $p < 0.001$), followed by Ng (AUC = 0.69, CI: 0.59–0.79, $p = 0.001$) and NPTX2 (AUC = 0.63, CI: 0.52–0.73, $p = 0.019$) to discriminate bvFTD from AD. GluR4 was not predictive between these two diagnostic groups. The AUC of NfL (AUC= 0.75, CI: 0.65–0.84, $p < 0.001$) was higher than all synaptic biomarkers except SNAP25. The selected biomarker panel using Wald's backward selection method, for differential diagnosis of bvFTD from AD, consisted of NfL, SNAP25, Ng, and GluR4 (AUC = 0.86, CI: 0.79–0.92, $p < 0.001$).

Associations of the CSF synaptic biomarkers and NfL with cognition and disease severity

The correlations of the CSF biomarkers with cognition and disease severity are shown in Fig. 3, Supplementary Figure 1, and Supplementary Figure 2. There were no associations between any of the synaptic markers and social cognition (Ekman 60 faces test) or disease severity in bvFTD. Lower CSF concentrations of NPTX2 (Pearson's $r = 0.29$, $p = 0.036$) and GluR4 (Pearson's $r = 0.34$, $p = 0.014$) were weakly associated with worse

Table 4 The AUC of the measured biomarkers as determined using ROC analysis

Biomarker	bvFTD vs PPD				bvFTD vs AD			
	AUC	CI	Specificity (%)	Sensitivity (%)	AUC	CI	Specificity (%)	Sensitivity (%)
	NfL	0.95***	0.91–0.99	81.0	95.8	0.76***	0.66–.86	88.3
SNAP25	0.55	0.44–0.66	49.1	64.8	0.79***	0.70–0.87	81.7	68.4
Ng	0.53	0.43–0.64	53.0	52.1	0.69**	0.59–0.79	65.0	66.7
NPTX2	0.71***	0.62–0.81	70.2	66.2	0.63*	0.52–0.73	88.3	42.1
GluR4	0.56	0.45–0.67	70.4	46.5	0.56	0.45–0.66	44.0	68.5
Panel	0.96***	0.93–0.99	87.7	91.5	0.86***	0.79–0.92	98.2	61.1

Wald's backward logistic regression was used to select the panel of CSF biomarkers. The biomarker panels selected were as follows: bvFTD vs PPD–NfL, NPTX2; bvFTD vs AD– NfL, SNAP25, Ng, GluR4. AUCs are shown for biomarkers alone. bvFTD, behavioral variant frontotemporal dementia; PPD, primary psychiatric disorders; AD, Alzheimer's disease; AUC, area under curve; ROC, receiver operating characteristics; CI, confidence interval; NfL, neurofilament; SNAP25, synaptosomal-associated protein 25; Ng, neurogranin; NPTX2, neuronal pentraxin 2. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$

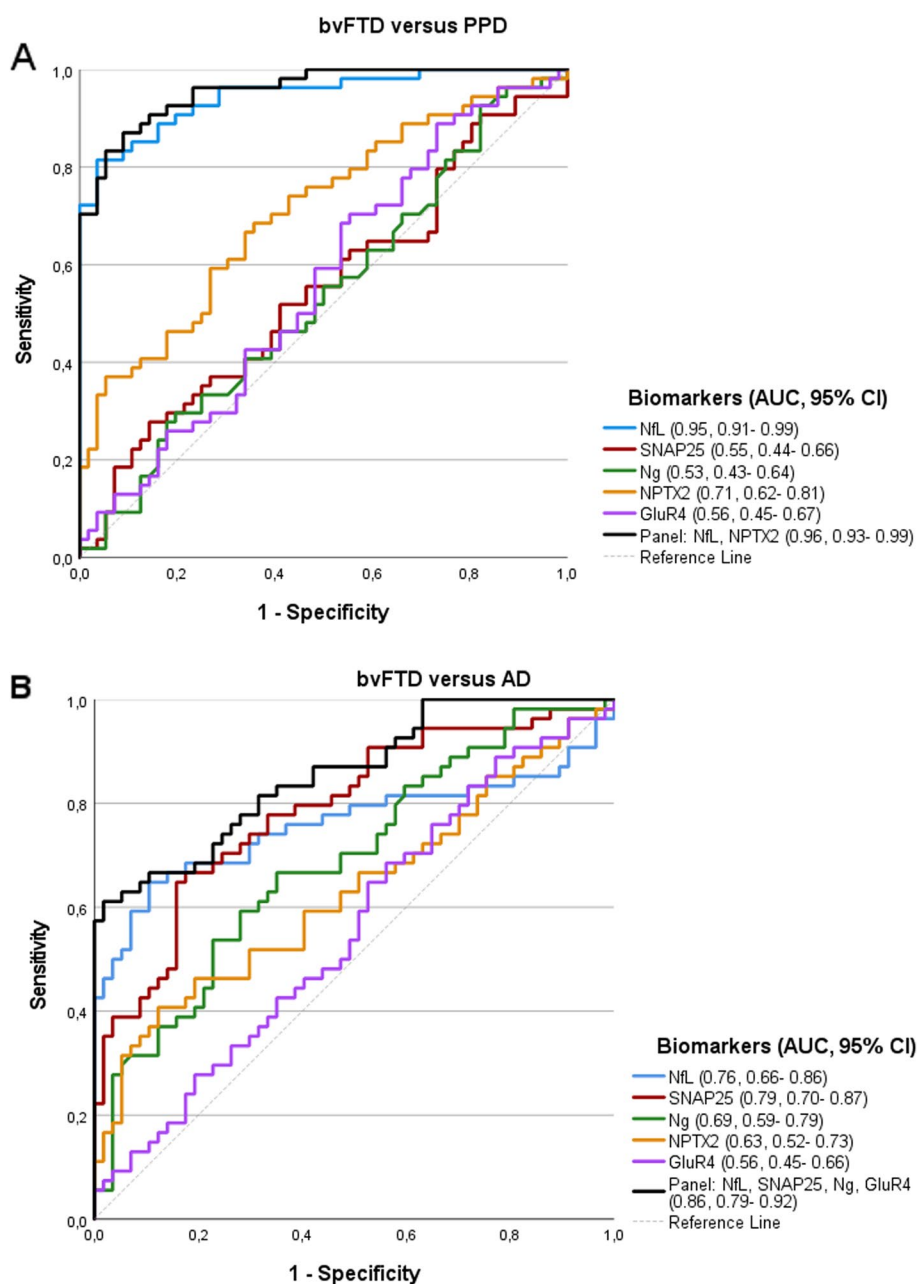


Fig. 2 ROC curves showing the differentiation accuracy between the diagnostic groups of biomarkers in CSF. **A** bvFTD versus PPD, Panel-NfL, NPTX2. **B** bvFTD versus AD, Panel-NfL, SNAP25, Ng, GluR4. AUCs are shown for biomarkers alone. bvFTD, behavioral variant frontotemporal dementia; PPD, primary psychiatric disorders; AD, Alzheimer's disease; AUC, area under curve; ROC, receiver operating characteristics; NfL, neurofilament; SNAP25, synaptosomal-associated protein 25; Ng, neurogranin; NPTX2, neuronal pentraxin 2; GluR4, glutamate receptor 4

performance of total cognitive score in bvFTD. Furthermore, lower GluR4 concentrations were also moderately associated with worse absolute MMSE scores in bvFTD (Pearson's $r = 0.41, p = 0.002$) and weakly associated with worse executive functioning (Pearson's $r = 0.36, p = 0.011$). In patients with AD, lower NPTX2 concentrations were moderately associated with worse performance

scores on the cognitive domain language (Pearson's $r = 0.43, p < 0.001$). Counterintuitively, in AD, higher CSF SNAP25 associated weakly with better performance on the domain attention (Pearson's $r = 0.30, p = 0.022$). No significant correlations were detected between the CSF biomarkers and cognitive scores in patients with PPD or in controls. In the bvFTD and PPD groups, there were no

Pearson's Partial Correlation	-1 -0.5 0 0.5 1																			
	bvFTD					PPD					AD					CONTROLS				
	NfL	SNAP25	Ng	NPTX2	GluR4	NfL	SNAP25	Ng	NPTX2	GluR4	NfL	SNAP25	Ng	NPTX2	GluR4	NfL	SNAP25	Ng	NPTX2	GluR4
Memory	-0.06	-0.03	0.25	-0.03	0.16	0.17	-0.03	0.14	-0.18	0.14	-0.17	-0.09	-0.12	0.13	0.06	0.26	0.12	0.02	0.01	0.01
Attention	0.05	0.15	0.13	0.18	0.25	0.06	-0.01	-0.19	0.12	0.12	0.02	0.30*	0.29	0.16	0.16	0.22	0.14	0.19	0.19	-0.26
Executive functioning	0.05	0.05	0.09	0.15	0.36*	0.10	-0.16	-0.08	-0.03	0.02	-0.17	0.11	0.16	0.08	0.13	0.30	0.31	-0.15	0.17	-0.12
Language	-0.12	-0.09	0.06	0.08	-0.14	0.14	0.06	0.09	0.09	0.07	0.03	0.13	0.14	0.43***	0.07	0.05	-0.04	0.13	0.04	-0.32
Visuospatial functioning	-0.13	0.32	0.14	-0.08	0.10	0.24	0.23	0.23	0.20	0.11	-0.06	0.04	-0.02	0.00	-0.20	-0.29	0.18	-0.01	0.14	0.22
Total Cognitive Score	-0.01	0.19	0.17	0.29*	0.34*	0.13	0.03	-0.01	0.13	0.18	-0.02	0.19	0.19	0.21	0.23	0.21	0.18	0.04	0.17	-0.17
MMSE	-0.14	0.09	-0.06	0.22	0.41**	0.04	0.00	0.19	0.07	0.18	-0.20	-0.05	-0.07	0.07	0.11	-0.02	-0.10	-0.12	-0.01	-0.12
Mood	0.13	-0.16	0.03	-0.14	-0.32	0.00	-0.19	-0.17	-0.04	-0.23	-0.32	-0.13	-0.03	-0.18	-0.22	-0.26	-0.22	-0.20	-0.29	0.00
Ekman Faces Test	-0.13	-0.17	-0.38	0.18	-0.01	-0.02	-0.12	0.09	-0.14	0.01										
FTLD-CDR	-0.02	0.10	0.15	-0.07	-0.10															

Fig. 3 Correlation matrix of the fluid biomarkers to cognitive test performance and social test scores in patients with bvFTD, PPD, AD, and controls. The associations are shown as Pearson's partial correlations, controlling for age. bvFTD, behavioral variant frontotemporal dementia; PPD, primary psychiatric disorders; AD, Alzheimer's disease; NfL, neurofilament light; SNAP25, synaptosomal-associated protein 25; Ng, neurogranin; NPTX2, neuronal pentraxin 2; GluR4, glutamate receptor 4; MMSE, mini-mental state examination; FTLD-CDR, frontotemporal lobe dementia-cognitive dementia rating. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$

significant correlations between cognitive scores and the generated biomarker panels (bvFTD_PPD, bvFTD_AD, Supplementary Figure 1). In the AD group, the panel bvFTD_AD correlated weakly with attention (Pearson's $r = -0.30$, $p = 0.026$), and in controls, the panel bvFTD_PPD correlated moderately with visuospatial functioning (Pearson's $r = -0.42$, $p = 0.012$).

Discussion

In this study, we found differential concentrations of CSF synaptic markers between bvFTD, PPD, AD, and controls in which reduced NPTX2 concentrations were bvFTD specific, and increased concentrations of SNAP25 and Ng were AD specific. Adding NPTX2 to NfL in the biomarker panel to distinguish bvFTD and PPD patients provided added diagnostic value, although limited. Synaptic biomarker concentrations did not correlate with social cognition nor disease severity in FTD and showed weak correlations with cognitive performance. These results indicate that NPTX2, alongside NfL, may provide further insights into bvFTD pathophysiology, although it is relatively less suitable clinically as a diagnostic biomarker.

NfL is a reliable biomarker of neuroaxonal damage and is used for the diagnosis (although limited due to its non-disease specificity), prognosis, and monitoring of treatment response in several neurodegenerative conditions [38]. Several clinical reports have asserted the association of NfL with grey matter and hippocampal atrophy, neuronal impairment, and loss of cognition in neurodegenerative dementias [17–20]. Accumulating clinical evidence further suggests that NfL is a promising biomarker in clinical settings to differentiate patients with bvFTD from PPD [39]. However, high levels of NfL are not specific to bvFTD as this biomarker can be strongly elevated in several other neurodegenerative conditions as well [2, 40, 41]. Furthermore, clinical reports suggest that synaptic

dysfunction precedes atrophy in patients with FTD [11, 14], which highlights the need for novel biomarkers that may aid in earlier and more accurate diagnosis of bvFTD over PPD.

Our findings of lower NPTX2 concentrations in sporadic FTD compared with PPD and controls are in line with our previous pilot study and with a recent study among patients with genetic forms of FTD, in which symptomatic mutation carriers showed lower concentrations of CSF NPTX2 compared with controls [13]. NPTX2 has also recently been identified as a promising biomarker for progression in genetic FTD [13]. NPTX2 is involved in the formation and stabilization of synapses, facilitating proper communication between neurons in the brain, and plays a crucial role in synaptic function and plasticity [42]. Studies have shown decreased synaptic density in the brains of FTD patients, particularly in brain regions affected by the disease including the salient network, inducing the characteristic impaired social cognition that is a hallmark feature of FTD [11]. Experimental evidence further suggests that downregulation of NPTX2 may lead to increased complement-mediated microglial activation, thereby causing abnormal elimination of synapses [43]. The downregulation of NPTX2 in both genetic and sporadic bvFTD may thus reflect a shared pathophysiology within the FTD disease heterogeneity and suggest that NPTX2 may play a crucial role in the pathogenesis of FTD by contributing to synaptic dysfunction [42]. Further investigation of NPTX2 and its mechanisms in FTD could provide valuable insights into the disease mechanisms and potentially lead to the development of novel therapeutic strategies targeting synaptic dysfunction.

While it has been reported elsewhere that CSF concentration of GluR4 is decreased in patients with mood disorder and schizophrenia compared with healthy controls [16], we did not find any diagnostic significance of this

biomarker in this cohort. This might be due to our heterogeneous sample of PPD, including various subtypes such as mood disorders, personality disorders, autism spectrum disorder, anxiety disorder, functional disorder, schizophrenia, and other psychiatry, of which individual group levels did not reach statistically significant thresholds of lower GluR4.

Synaptic pathology is a shared mechanism across diseases, yet evidence presented here and elsewhere indicates that synaptic proteins participate differentially in various disease pathogeneses, underscoring the distinct impairments in synaptic functionality across diseases. For example, the AD-specific increase of CSF SNAP25 and Ng compared with bvFTD that we reported, corroborates previous findings [44, 45]. Ng is a postsynaptic protein that is important for maintaining synaptic plasticity and regulating calcium ion influxes, while SNAP25 is a presynaptic protein that plays a crucial role in synaptic vesicle fusion and neurotransmitter release, and both these proteins have been shown to play a key role in AD disease pathophysiology, although they may be less clinically relevant for FTD [10, 45–48]. While brain regions affected in AD, i.e., the hippocampus and cortex have a high expression of Ng, the anatomical distribution of SNAP25 is not well known, although it is expressed in the cortex [49–52]. Thus, a possible hypothesis for the increased concentrations of synaptic proteins SNAP25 and Ng in AD but not in bvFTD could be due to the topography of brain atrophy they reflect [45].

In our cohort, bvFTD patients performed worse on social cognition testing compared with PPD, but no association was found with CSF synaptic markers which might be due to the limited test scores available per diagnostic group, i.e., only 13 for bvFTD and 9 for PPD. A previous study, using [¹⁸F]UCBH-PET as a tracer for synaptic vesicle protein 2A (SV2A) which reflects synaptic density, showed a trend for synaptic loss in the temporal social brain in bvFTD, highlighting the clinical relevance of synaptopathy in disease pathophysiology of FTD [14]. Further studies assessing CSF synaptic markers might elucidate if SV2A is a superior synaptic marker in CSF correlating with social cognition in larger patient groups.

The association of the synaptic biomarkers with other cognitive functioning was absent or only moderate to weak in bvFTD and AD, while there were no correlations found in patients with PPD or with controls. Moreover, we did not detect any association of the synaptic proteins or NfL with FTLN-CDR disease severity scores [53, 54]. One plausible reason could be that the cohort included a homogenous sample of bvFTD patients with mild to moderate bvFTD disease severity (FTLN-CDR \leq 16). The direction of correlation detected in the AD group between SNAP25 and attention was counterintuitive, and

thus follow-up studies with greater statistical power are necessary to evaluate these findings.

The strengths of this study lie in including the assessment of concentrations of CSF synaptic proteins involved in several synaptic functions both upstream and downstream of the synapse, providing insight into pathophysiological mechanisms. Additionally, we included a well-phenotyped diverse PPD sample as a comparative group, which is the most important and clinically challenging to differentially diagnose from bvFTD, resembling clinical practice. There are also some limitations. For example, the odds ratios of the synaptic biomarkers for diagnostic distinction of bvFTD versus PPD and AD were modest and the clinical relevance of these biomarkers, particularly NPTX2, demands to be evaluated in larger cohorts. Since the PPD sample was heterogeneous, disease-specific synaptic concentrations in PPD could not be assessed and should be included in future studies. Furthermore, the sample sizes for cognitive test scores were limited, such as for social cognition (Ekman 60 faces test), disease severity (FTLN-CDR), and the domain visuospatial functioning.

Conclusions

We conclude that synaptic biomarker NPTX2 has additional, although limited diagnostic value to NfL in the differential diagnosis of bvFTD versus PPD. Our findings contribute insight into disease-specific mechanisms in bvFTD, by showing the bvFTD-specific decrease in concentrations of NPTX2. Furthermore, given that NfL likely reflects neuronal atrophy [17, 18], it is a clinically relevant biomarker at a progressed stage of the disease. Further investigation of NPTX2 and its mechanisms in bvFTD could provide valuable insights into the disease mechanisms for early diagnosis and prognosis, as well as potentially lead to the development of novel therapeutic strategies.

Abbreviations

AD	Alzheimer's disease
ANCOVA	Analysis of covariance
ANOVA	Analysis of variance
AUC	Area under curve
bvFTD	Behavioral variant frontotemporal dementia
CSF	Cerebrospinal fluid
CV	Coefficient of variation
FAB	Frontal assessment battery
FTD	Frontotemporal dementia
FTLN-CDR	Frontotemporal lobar degeneration-modified clinical dementia rating
GDS	Geriatric depression scale
GluR4	Glutamate receptor 4
MMSE	Mini-mental state examination
NfL	Neurofilament light
Ng	Neurogranin
NPTX2	Neuronal pentraxin 2
OR	Odds ratio

PPD	Primary psychiatric disorders
RAVLT	Rey auditory verbal learning test
ROC	Receiver operating characteristics
SNAP25	Synaptosomal-associated protein of 25 kDa
TMT	Trail making test
VAT	Visual association test
VOSP	Visual object and space perception

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s13195-024-01409-8>.

Additional file 1: Supplementary Figure 1. Correlation matrix of the fluid biomarkers to cognitive test performance and social test scores in patients with bvFTD, PPD, AD, and controls. The associations are shown as Pearson's partial correlations, controlling for age. bvFTD: behavioral variant frontotemporal dementia, PPD: primary psychiatric disorders, AD: Alzheimer's disease, NfL: neurofilament light, SNAP25: synaptosomal associated protein 25, Ng: neurogranin, NPTX2: neuronal pentraxin 2, GluR4: Glutamate receptor 4, MMSE: mini-mental state examination, FTLD-CDR: frontotemporal lobe dementia- cognitive dementia rating. Panel bvFTD_PPD: NfL, NPTX2, Panel bvFTD_AD: NfL, SNAP25, Ng, GluR4 (both differential diagnostic panels selected using backward logistic regression models). * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$

Additional file 2: Supplementary Figure 2. Visualization of correlations between biomarkers and cognitive test scores. A) bvFTD: NPTX2 versus total cognitive score, B) bvFTD: GluR4 versus total cognitive score, C) bvFTD: GluR4 versus executive functioning, D) bvFTD: GluR4 versus MMSE scores, E) AD: NPTX2 versus language and F) AD: SNAP25 versus attention. bvFTD: behavioral variant frontotemporal dementia, AD: Alzheimer's disease, SNAP25: synaptosomal associated protein 25, NPTX2: neuronal pentraxin 2, GluR4: Glutamate receptor 4, MMSE: mini-mental state examination.

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Authors' contributions

S.D. performed development and analytical validations of the immunoassays, clinical sample measurements, data analysis, and figure preparation, and wrote the first draft of the manuscript. M.P.E.V.E. performed clinical cohort design and sample selection, clinical assessments of the patients and procured clinical data, data analysis, and wrote the first draft of the manuscript. J.G. performed development and analytical validations of the immunoassays and reviewed and critiqued the manuscript. D.J. performed development and analytical validations of the immunoassays. B.B. performed analytical validations of the immunoassays, clinical sample measurements, and reviewed the manuscript. J.L.P.F. provided critical information regarding clinical neuropsychological evaluations and reviewed the manuscript. Y.A.L.P. was responsible for project supervision, clinical cohort design, and reviewed the manuscript. C.E.T. was responsible for project supervision, clinical cohort design, and reviewed the manuscript. E.V. was responsible for project supervision, clinical cohort design, and reviewed the manuscript. I.M.W.V. was responsible for project supervision, provided assistance with data analysis techniques, as well as extensive critical review of the manuscript. S.D. and M.P.E.V.E. have contributed equally to this manuscript. All authors have approved the submission of this manuscript and consent to its publication. All authors reviewed the manuscript.

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Availability of data and materials

Anonymized data can be made available upon reasonable request and consultation with the involved authorities. Additional datasets generated during and/or analyzed during the current study are available from the corresponding author upon reasonable request.

Declarations

Ethics approval and consent to participate

All clinical CSF samples were requested via the Amsterdam Dementia Cohort biobank. The study was approved by the Medical Ethical Committee of Amsterdam University Medical Center. All participants provided informed consent and the study has been carried out by the Declaration of Helsinki.

Consent for publication

Not applicable.

Competing interests

S.D., and J.G., are employees of ADx NeuroSciences, Gent, Belgium. E.V. is the co-founder of ADx NeuroSciences. D.J. is a former employee of ADx NeuroSciences, Gent, Belgium, and is recently retired. C.E.T. has a collaboration contract with ADx NeuroSciences, Quantex, and Eli Lilly, and has performed contract research or received grants from AC-Immune, Axon Neurosciences, Bioconnect, Bioorchestra, Brainstorm Therapeutics, Celgene, EIP Pharma, Eisai, Grifols, Novo Nordisk, PeopleBio, Roche, Toyama, and Vivoryon. She serves on editorial boards of *Medicard Neurologie/Springer*, *Alzheimer Research and Therapy*, *Neurology: Neuroimmunology & Neuroinflammation*, and is the editor of a *Neuroinformatics* book Springer. All the other authors declare that they have no competing interests.

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References

- Onyike CU, Diehl-Schmid J. The epidemiology of frontotemporal dementia. *Int Rev Psychiatry*. 2013;25(2):130–7.
- Del Campo M, Zetterberg H, Gandy S, Onyike CU, Oliveira F, Udeh-Momoh C, et al. New developments of biofluid-based biomarkers for routine diagnosis and disease trajectories in frontotemporal dementia. *Alzheimers Dement*. 2022;18(11):2292–307.
- Ducharme S, Dols A, Laforce R, Devenney E, Kumfor F, van den Stock J, et al. Recommendations to distinguish behavioural variant frontotemporal dementia from psychiatric disorders. *Brain*. 2020;143(6):1632–50.
- Woolley JD, Khan BK, Murthy NK, Miller BL, Rankin KP. The diagnostic challenge of psychiatric symptoms in neurodegenerative disease; rates of and risk factors for prior psychiatric diagnosis in patients with early neurodegenerative disease. *The Journal of clinical psychiatry*. 2011;72(2):126.

5. Vijverberg EG, Wattjes MP, Dols A, Krudop WA, Möller C, Peters A, et al. Diagnostic accuracy of MRI and additional [18F] FDG-PET for behavioral variant frontotemporal dementia in patients with late onset behavioral changes. *Journal of Alzheimer's Disease*. 2016;53(4):1287–97.
6. Henstridge CM, Sideris DI, Carroll E, Rotariu S, Salomonsson S, Tzioras M, et al. Synapse loss in the prefrontal cortex is associated with cognitive decline in amyotrophic lateral sclerosis. *Acta neuropathologica*. 2018;135(2):213–26.
7. Mackenzie IR, Neumann M. Molecular neuropathology of frontotemporal dementia: insights into disease mechanisms from postmortem studies. *Journal of neurochemistry*. 2016;138:54–70.
8. Lashley T, Rohrer JD, Mead S, Revesz T. An update on clinical, genetic and pathological aspects of frontotemporal lobar degenerations. *Neuropathology and applied neurobiology*. 2015;41(7):858–81.
9. Rademakers R, Neumann M, Mackenzie IR. Advances in understanding the molecular basis of frontotemporal dementia. *Nature Reviews Neurology*. 2012;8(8):423–34.
10. Cervantes González A, Irwin DJ, Alcolea D, McMillan CT, Chen-Plotkin A, Wolk D, et al. Multimarker synaptic protein cerebrospinal fluid panels reflect TDP-43 pathology and cognitive performance in a pathological cohort of frontotemporal lobar degeneration. *Molecular Neurodegeneration*. 2022;17(1):1–12.
11. Salmon E, Bahri MA, Plenevaux A, Becker G, Seret A, Delhay E, et al. In vivo exploration of synaptic projections in frontotemporal dementia. *Scientific reports*. 2021;11(1):1–10.
12. Sogorb-Esteve A, Nilsson J, Swift IJ, Heller C, Bocchetta M, Russell LL, et al. Differential impairment of cerebrospinal fluid synaptic biomarkers in the genetic forms of frontotemporal dementia. *Alzheimer's research & therapy*. 2022;14(1):1–12.
13. Van Der Ende EL, Xiao M, Xu D, Poos JM, Panman JL, Jiskoot LC, et al. Neuronal pentraxin 2: a synapse-derived CSF biomarker in genetic frontotemporal dementia. *Journal of Neurology, Neurosurgery & Psychiatry*. 2020;91(6):612–21.
14. Malpetti M, Jones PS, Cope TE, Holland N, Naessens M, Rouse MA, et al. Synaptic loss in frontotemporal dementia revealed by [11C] UCB-J positron emission tomography. *Ann Neurol*. 2023;93(1):142–54. <https://doi.org/10.1002/ana.26543>.
15. Das S, Goossens J, Jacobs D, Dewit N, Pijnenburg YAL, In 't Veld S, et al. Synaptic biomarkers in the cerebrospinal fluid associate differentially with classical neuronal biomarkers in patients with Alzheimer's disease and frontotemporal dementia. *Alzheimers Res Ther*. 2023;15(1):62.
16. Gómez de San José N, Goossens J, Al Shweiki MR, Halbgebauer S, Oeckl P, Steinacker P, et al. Glutamate receptor 4 as a fluid biomarker for the diagnosis of psychiatric disorders. *J Psychiatr Res*. 2022;156:390–97. <https://doi.org/10.1016/j.jpsychires.2022.10.010>.
17. Dhiman K, Gupta VB, Villemagne VL, Eratne D, Graham PL, Fowler C, et al. Cerebrospinal fluid neurofilament light concentration predicts brain atrophy and cognition in Alzheimer's disease. *Alzheimers Dement (Amst)*. 2020;12(1):e12005. <https://doi.org/10.1002/dad2.12005>.
18. Kartau M, Melkas S, Kartau J, Arola A, Laakso H, Pitkänen J, et al. Neurofilament light level correlates with brain atrophy, and cognitive and motor performance. *Front Aging Neurosci*. 2022;14: 939155.
19. Walia N, Eratne D, Loi SM, Farrand S, Li Q-X, Malpas CB, et al. Cerebrospinal fluid neurofilament light and cerebral atrophy in younger-onset dementia and primary psychiatric disorders. *Intern Med J*. 2023;53(9):1564–69. <https://doi.org/10.1111/imj.15956>.
20. Mattsson N, Cullen NC, Andreasson U, Zetterberg H, Blennow K. Association Between longitudinal plasma neurofilament light and neurodegeneration in patients with Alzheimer disease. *JAMA Neurol*. 2019;76(7):791–9.
21. Verde F, Otto M, Silani V. Neurofilament light chain as biomarker for amyotrophic lateral sclerosis and frontotemporal dementia. *Front Neurosci*. 2021;15:679199. <https://doi.org/10.3389/fnins.2021.679199>.
22. Katsiko K, Cajanus A, Jääskeläinen O, Kontkanen A, Hartikainen P, Korhonen VE, et al. Serum neurofilament light chain is a discriminative biomarker between frontotemporal lobar degeneration and primary psychiatric disorders. *J Neurol*. 2020;267(1):162–7.
23. Varhaug KN, Torkildsen Ø, Myhr KM, Vedeler CA. Neurofilament light chain as a biomarker in multiple sclerosis. *Front Neurol*. 2019;10:338. <https://doi.org/10.3389/fneur.2019.00338>.
24. Khalil M, Teunissen CE, Otto M, Piehl F, Sormani MP, Gatteringer T, et al. Neurofilaments as biomarkers in neurological disorders. *Nature Reviews Neurology*: Nature Publishing Group; 2018. p. 577–89.
25. Molinuevo JL, Rabin LA, Amarioglio R, Buckley R, Dubois B, Ellis KA, et al. Implementation of subjective cognitive decline criteria in research studies. *Alzheimer's & Dementia*. 2017;13(3):296–311.
26. van der Flier WM, Pijnenburg YA, Prins N, Lemstra AW, Bouwman FH, Teunissen CE, et al. Optimizing patient care and research: the Amsterdam Dementia Cohort. *Journal of Alzheimer's disease*. 2014;41(1):313–27.
27. Van Der Flier WM, Scheltens P. Amsterdam dementia cohort: performing research to optimize care. *Journal of Alzheimer's Disease*. 2018;62(3):1091–111.
28. Groot C, van Loenhoud AC, Barkhof V, van Berckel BNM, Koene T, Teunissen CC, et al. Differential effects of cognitive reserve and brain reserve on cognition in Alzheimer disease. *Neurology*. 2018;90(2): e149.
29. Van Der Flier WM, Scheltens P. Amsterdam dementia cohort: performing research to optimize care. *Alzheimers Dis*. 2018;62(3):1091–1111. <https://doi.org/10.3233/JAD-170850>.
30. Willems EAJ, van Maurik IS, Tijms BM, Bouwman FH, Franke A, Hubeek I, et al. Diagnostic performance of Elecsys immunoassays for cerebrospinal fluid Alzheimer's disease biomarkers in a nonacademic, multicenter memory clinic cohort: the ABIDE project. *Alzheimer's and Dementia: Diagnosis, Assessment and Disease Monitoring*. 2018;10:563–72.
31. McKhann GM, Knopman DS, Chertkow H, Hyman BT, Jack CR Jr, Kawas CH, et al. The diagnosis of dementia due to Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimers Dement*. 2011;7(3):263–9.
32. Rascovsky K, Hodges JR, Knopman D, Mendez MF, Kramer JH, Neuhaus J, et al. Sensitivity of revised diagnostic criteria for the behavioural variant of frontotemporal dementia. *Brain*. 2011;134(Pt 9):2456–77.
33. American Psychiatric Association, DSM-5 Task Force. *Diagnostic and statistical manual of mental disorders: DSM-5™* (5th ed.). American Psychiatric Publishing, Inc.; 2013. <https://doi.org/10.1176/appi.books.9780890425596>.
34. Knopman DS, Kramer JH, Boeve BF, Caselli RJ, Graff-Radford NR, Mendez MF, et al. Development of methodology for conducting clinical trials in frontotemporal lobar degeneration. *Brain*. 2008;131(11):2957–68.
35. Das S, Dewit N, Jacobs D, Pijnenburg YAL, In 't Veld SGJG, Coppens S, et al. A novel neurofilament light chain ELISA validated in patients with Alzheimer's disease, frontotemporal dementia, and subjective cognitive decline, and the evaluation of candidate proteins for immunoassay calibration. *Intern J Mol Sci*. 2022;23(13):7221.
36. Bolsewig K, Hok-A-Hin YS, Sepe FN, Boonkamp L, Jacobs D, Bellomo G, et al. A combination of neurofilament light, glial fibrillary acidic protein, and neuronal pentraxin-2 discriminates between frontotemporal dementia and other dementias. *Journal of Alzheimer's Disease*. 2022;90:363–80.
37. Andreasson U, Perret-Liaudet A, van Waalwijk van Doorn LJC, Blennow K, Chiasserini D, Engelborghs S, et al. A practical guide to immunoassay method validation. *Front Neurol*. 2015;19(6):179.
38. Gaetani L, Blennow K, Calabresi P, Di Filippo M, Parnetti L, Zetterberg H. Neurofilament light chain as a biomarker in neurological disorders. *J Neurol, Neurosurg Psychiatry*: BMJ Publishing Group. 2019;90(8):870–81.
39. Eratne D, Loi SM, Li QX, Stehmann C, Malpas CB, Santillo A, et al. Cerebrospinal fluid neurofilament light chain differentiates primary psychiatric disorders from rapidly progressive, Alzheimer's disease and frontotemporal disorders in clinical settings. *Alzheimer's Dement*. 2022;18(11):2218–33.
40. Ulugut H, Pijnenburg YAL. Frontotemporal dementia: past, present, and future. *Alzheimers Dement*. 2023;19(11):5253–63.
41. Illán-Gala I, Lleo A, Karydas A, Staffaroni AM, Zetterberg H, Sivasankaran R, et al. Plasma tau and neurofilament light in frontotemporal lobar degeneration and alzheimer disease. *Neurology*. 2021;96(5):e671–83.
42. Gómez de San José N, Massa F, Halbgebauer S, Oeckl P, Steinacker P, Otto M. Neuronal pentraxins as biomarkers of synaptic activity: from physiological functions to pathological changes in neurodegeneration. *J Neural Transm (Vienna)*. 2022;129(2):207–30. <https://doi.org/10.1007/s00702-021-02411-2>.
43. Zhou J, Wade SD, Graykowski D, Xiao M-F, Zhao B, Giannini LAA, et al. The neuronal pentraxin Nptx2 regulates complement activity and restrains microglia-mediated synapse loss in neurodegeneration. *Science Translational Medicine*. 2023;15(689):eadf0141.

44. Kivisäkk P, Carlyle BC, Sweeney T, Quinn JP, Ramirez CE, Trombetta BA, et al. Increased levels of the synaptic proteins PSD-95, SNAP-25, and neurogranin in the cerebrospinal fluid of patients with Alzheimer's disease. *Alzheimer's Research & Therapy*. 2022;14(1):58.
45. Clarke MTM, Brinkmalm A, Foiani MS, Woollacott IOC, Heller C, Heslegrave A, et al. CSF synaptic protein concentrations are raised in those with atypical Alzheimer's disease but not frontotemporal dementia. *Alzheimer's Res Ther*. 2019;11(1):105.
46. Wang C, Tu J, Zhang S, Cai B, Liu Z, Hou S, et al. Different regions of synaptic vesicle membrane regulate VAMP2 conformation for the SNARE assembly. *Nature Communications*. 2020;11(1):1531.
47. Goetzl EJ, Kapogiannis D, Schwartz JB, Lobach IV, Goetzl L, Abner EL, et al. Decreased synaptic proteins in neuronal exosomes of frontotemporal dementia and Alzheimer's disease. *FASEB Journal*. 2016;30(12):4141–8.
48. Brinkmalm A, Brinkmalm G, Honer WG, Frölich L, Hausner L, Minthon L, et al. SNAP-25 is a promising novel cerebrospinal fluid biomarker for synapse degeneration in Alzheimer's disease. *Mol Neurodegener*. 2014;9:53.
49. Portelius E, Olsson B, Höglund K, Cullen NC, Kvartsberg H, Andreasson U, et al. Cerebrospinal fluid neurogranin concentration in neurodegeneration: relation to clinical phenotypes and neuropathology. *Acta Neuropathologica*. 2018;136(3):363–76.
50. Represa A, Deloulme JC, Sensenbrenner M, Ben-Ari Y, Baudier J. Neurogranin: immunocytochemical localization of a brain-specific protein kinase C substrate. *J Neurosci*. 1990;10(12):3782–92.
51. Yamamori S, Itakura M, Sugaya D, Katsumata O, Sakagami H, Takahashi M. Differential expression of SNAP-25 family proteins in the mouse brain. *Journal of Comparative Neurology*. 2011;519(5):916–32.
52. Glavan G, Schliebs R, Živin M. Synaptotagmins in neurodegeneration. *The Anatomical Record*. 2009;292(12):1849–62.
53. Diehl-Schmid J, Pohl C, Ruprecht C, Wagenpfeil S, Foerstl H, Kurz A. The Ekman 60 Faces Test as a diagnostic instrument in frontotemporal dementia. *Archives of Clinical Neuropsychology*. 2007;22(4):459–64.
54. Research Australia N, Piguet AO, Hornberger M, Mioshi E, Hodges Fmed-Sci JR, Hodges J, et al. Behavioural-variant frontotemporal dementia: diagnosis, clinical staging, and management. *Lancet Neurol*. 2011;10:162–72.

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